

Histamine and Atopic Eczema

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Apart from increased production of immunoglobulin E antibodies and disturbed T-cell regulation, altered patterns of releasability of vasoactive mediators have been described in patients with atopic eczema. The best studied substance is histamine which is a classical inducer of pruritus in man. Elevated concentrations of histamine have been found *in vivo* in the skin and in the plasma of patients with atopic eczema especially during exacerbation of the disease. Similar findings have been described for other atopic diseases as extrinsic bronchial asthma. Histamine acts via characteristic receptors; symptoms as itch, wheal formation, mucus production, contraction of smooth muscle, tachycardia and hypotension are mediated via H₁-receptors, while H₂-effects include acid secretion in the stomach as well as the development of flush and itch reactions, blood pressure changes and cardiac arrhythmia. Of special interest is an inhibitory effect of histamine on lymphocyte reactions mediated via a H₂-receptor. The existence of a new H₃-receptor in the brain serving as autocrine feed-back inhibitor of histaminergic neurones has been established in the rat but not yet in man. *In vitro* an increased histamine releasability of peripheral leukocytes has been found after stimulation with a variety of different substances. The difference between patients with atopic eczema and normals is generally most pronounced after stimulation with anti-IgE. There is, however, a tendency towards an increased spontaneous histamine release compared to normals. The release reaction of histamine seems to occur more rapidly in atopics compared to normals. Among possible factors influencing histamine releasability the imbalance in the cyclic nucleotide system (increased response of cGMP to cholinergic stimulation and decreased response of cAMP to β -adrenergic stimulation) might play a pathogenetic role. Arachidonic acid metabolites known to regulate histamine release (PGE₂ inhibits histamine release while cyclooxygenase blockers enhance histamine release and lipoxigenase blockers inhibits histamine release) also may be of relevance. Histamine definitely is not the only relevant mediator substance in the pathophysiology of atopic eczema; it may, however, serve as a marker of mast cell and basophil activation. Clinical trials with various antihistamines have shown some therapeutic benefit in the management of atopic eczema patients. Future studies in the field of mediator research may

lead to new therapeutic approaches for the treatment of atopic eczema.

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HISTAMINE EFFECTS

Although known for almost 80 years, histamine still remains a fascinating substance for many researchers (7, 8, 17, 18, 28, 46, 47, 53, 69, 83). The definition of the physiological role of histamine in health and disease remains incomplete. We know that histamine exerts powerful effects mainly via two receptors (Table I). The description of a new H₃-receptor in the brain deserves special interest (4, 81 a).

The role of histamine as a mediator of immediate-type hypersensitivity diseases (both allergic and pseudo-allergic in origin) is quite well established (7, 16, 17, 36, 68, 79). Similarly well defined is the H₂-mediated role of histamine in the induction of gastric acid secretion.

Recent interest has focussed on anti-inflammatory effects of histamine as a modulator of immune reactions acting predominantly on H₂-receptors on the surface of leukocytes there by inhibiting a variety of immune reactions (Table II) (9, 13, 72, 73, 81, 85).

In a study in 16 patients with atopic eczema we found an inhibitory effect of histamine upon poke-weed-mitogen(PWM)-induced lymphocyte stimulation (Fig. 1). This effect was shown to depend upon the presence of T8-lymphocytes in the cell suspension, as shown in Fig. 2: after depletion of T8-lymphocytes the inhibitory effect of histamine upon PWM-induced lymphocyte stimulation was no longer demonstrable in atopics nor in controls.

Role of histamine in the pathophysiology of atopic eczema

In spite of great progress in experimental and clinical allergology and dermatology in the last decades the

Table I. Histamine effects in human organs

Organ	Stimulatory	Inhibitory	None/negligible
<i>Vessels</i>			
Large veins (> 80 µm)	+	-	-
Arterioles, venules (20-30 µm)	-	+	-
Permeability of "capillaries" (postcapillary venules)	+	-	-
<i>Extravascular smooth muscle</i>			
Bronchi, gut	+	-	-
Uterus, bladder, gallbladder, iris	-	-	+
Stomach (secretion)	+	-	-
Salivary glands	+	-	-
<i>Heart</i>			
Rate, force, output	+	-	-
AV conduction	-	+	-
Ventricular arrhythmia	+	-	-
<i>Nervous system</i>			
Sensory fibers	+	-	-
Central effects	(+)?	-	-
<i>Endocrine system</i>			
Adrenal medulla	-	-	+
Leukocytes	-	+	-

pathogenesis of atopic eczema is still not well established.

Research interest has focussed on mainly three aspects:

- increased production of immunoglobulin E,
- disturbed T-cell regulation,
- altered pharmacological reactivity and releasability of vasoactive mediators (6, 11, 15, 23, 25, 26, 31, 32, 35, 41, 42, 43, 44, 48, 56, 57, 58, 62, 63, 80, 85, 94).

Previously we have advanced the concept of a "vicious cycle" of this different factors acting together in the pathophysiology of atopic eczema (61).

The best studied mediator substance is histamine which is a classical inducer of pruritus in man (57, 69). Elevated concentrations of histamine have been found in vivo in the skin and in the plasma of patients with atopic eczema especially during exacerbation of the disease (37, 38, 66, 74) (Table III). Similar findings have been described for other atopic diseases as extrinsic bronchial asthma (82). In vitro an increased histamine releasability of peripheral leukocytes has been found after stimulation with a variety of different substances by many other authors (5, 14, 15, 22,

51, 65, 69, 74, 90). The difference between patients with atopic eczema and normals is generally most pronounced after stimulation with anti-IgE. There is, however, a tendency towards an increased spontaneous histamine release compared to normals (69, 84). The release reaction of histamine seems to occur more rapidly in atopics compared to normals (91).

Among possible factors influencing histamine releasability the imbalance in the cyclic nucleotide system (increased response of cGMP to cholinergic stim-

Table II. Immune reactions inhibited by histamine

Mast cells and basophils (mediator release)
Neutrophil function (chemotaxis, phagocytosis, enzyme release)
Eosinophils
Macrophages (complement production)
Lymphocyte proliferation and lymphokine production
Lymphocyte cytotoxicity
Concanavalin A-induced suppressor cell activity

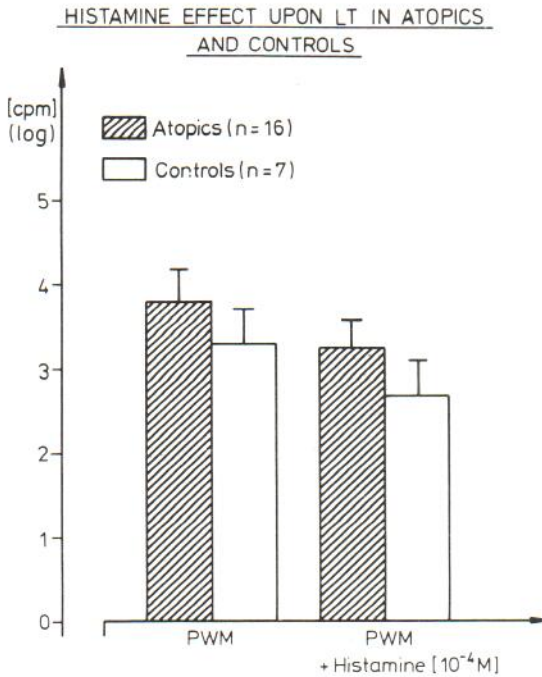


Fig. 1. Effect of histamine added to lymphocyte cultures upon lymphocyte transformation (LT) induced by pokeweed mitogen (PWM) in patients with atopic eczema and controls.

ulation and decreased response of cAMP to β -adrenergic stimulation) (1, 3, 12, 14, 21, 27, 34, 40, 43, 49, 54, 59, 64, 65, 78, 86, 87, 88) might play a pathogenetic role.

Arachidonic acid metabolites known to regulate histamine release (PGE₂ inhibits histamine release while cyclooxygenase blockers enhance histamine release and lipoxygenase blockers inhibits histamine release) (2, 16, 18, 19, 20, 50, 69, 91) also may be of relevance.

Histamine definitely is not the only relevant mediator substance in the pathophysiology of atopic eczema; it may, however, serve as a marker of mast cell and basophil activation. Similar results of increased formation of leukotriene B₄ in involved skin of patients with atopic eczema (75) as well as enhanced in vitro leukotriene B₄ secretion from peripheral leukocytes in atopic patients have been reported (76).

Psychosomatic interactions, histamine and atopic eczema

The in vivo and in vitro demonstrable dysregulation of the autonomic nervous system in patients with atopic eczema (33, 43, 44, 56, 68, 94) together with

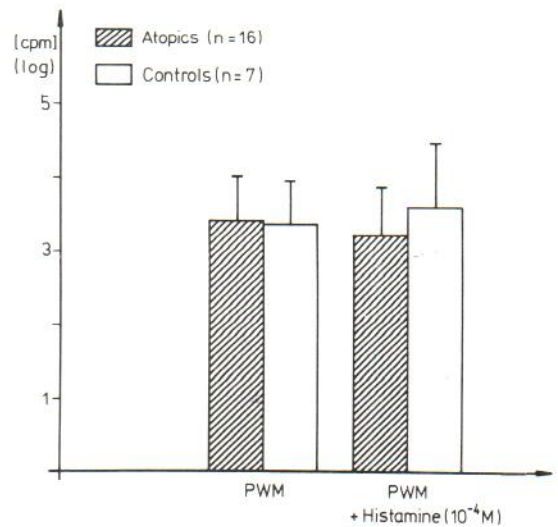


Fig. 2. Effect of histamine added to lymphocyte cultures upon lymphocyte transformation (LT) induced by pokeweed mitogen (PWM) in patients with atopic eczema and controls after specific depletion of T8-cells by rosetting technique with monoclonal antibodies (see ...)

the role of autonomic nervous system transmitters in regulating histamine release (inhibition by β -adrenergic, enhancement by cholinergic stimuli) (27, 46, 64, 65, 78, 86) and the possible existence of a newly described H₃-receptor in the brain might open a new field of research in order to more clearly define the nature of psychosomatic interactions in this disease (10, 67, 92). This new H₃-receptor has been demonstrated in the rat brain where it serves as an autocrine feed-back inhibitor of histaminergic neurones leading

Table III. *Histamine and atopic eczema*

In vivo

Histamine concentration elevated in the skin (\pm)
 Histamine elevated in plasma (during exacerbation)

In vitro

Altered releasability
 Increased release from basophil leukocytes
 Increased speed of release
 Influence of autonomic nervous system transmitters
 Modulation by arachidonic acid metabolites

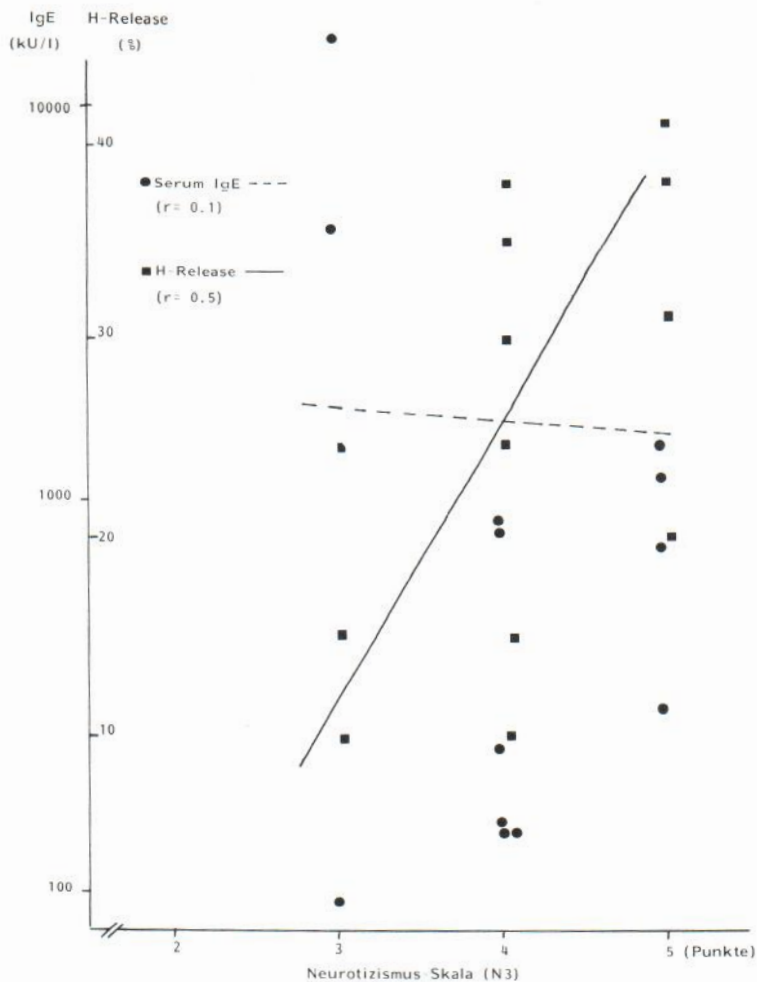


Fig. 3. Correlation of psychodiagnostic test results in the neuroticism scale of the "Hamburg Neuroticism and Extraversion Scale" (HANES) for children and adolescents to somatic findings as serum IgE concentration and in vitro histamine releasability after anti-IgE-stimulation of peripheral leukocytes.

to diminished histamine synthesis in and synaptic secretion from these cells (4, 81 a).

It has been shown by various authors that histamine release can be induced by stress in various forms (60, 69).

In a psychosomatic investigation using several psychodiagnostic tests in children with atopic eczema and control children with non-inflammatory dermatologic diseases we compared the results of the "Hamburg Neuroticism and Extraversion Scale" (HANES) for children and adolescents with somatic findings as extent of skin lesions, serum IgE-level and in vitro histamine releasability (67). As shown in Fig. 3 there was no correlation between the results of the psychodiagnostic tests to any somatic finding except for the slightly pronounced positive correlation between in vitro histamine releasability towards anti-IgE and neuroticism as measured in the HANES scale (Fig. 3).

Therapeutic approaches

Therapeutic approaches to histamine-mediated diseases can act at different stages (Table IV) from the inhibition of histamine synthesis via blockade of histamine release at different levels until specific antagonism of histamine effects (Table IV) (11, 24, 58, 68, 94).

In antihistamine therapy new developments include the production of non-sedating H1-antagonists, the combination of H1- and H2-antagonists as well as the application of H1-antagonists with mast cell blocking activity (Table V).

The side effects of classical H1-antagonists include mostly sedative effects; it seems of interest, however, that in double-blind studies these sedating side effects are observed regularly, especially in a certain percentage of atopic patients even under placebo (Table VI).

There is some controversy regarding the effect of

Table IV. Therapeutic modalities in histamine-mediated diseases

Histamine synthesis inhibitors
Histamine release blockers
cAMP-active substances
Flavonoids
Ca-antagonists (?)
Inosiplex (?)
Lipoxygenase inhibitors
Histamine antagonists

Table V. Antihistamine therapy

H1-antagonists (classic)
H1-antagonists (non-sedating)
H1-antagonists with mast cell blocking activity
H2-antagonists
H1- + H2-antagonists combined
H3-agonists or antagonists (?)

Table VI. "Antihistamine"-side effects under placebo treatment (%)

Sedation	13
Headache	6
Dryness of mouth	5
Diarrhoea	2
Nausea	2
Abdominal pain	2
CNS-stimulation	1
Exanthema	1
Increase in body weight	1

antihistamine therapy in atopic eczema. Especially with regard to the question whether sedating side-effects are essential for a possible therapeutic effect of antihistamines. In Table VII some studies from the literature are enlisted dealing with the efficacy of antihistamines upon itch or atopic eczema.

Obviously more studies will have to be done to really answer these questions. So far it seems to be fair to state that antihistamines can never represent the one and only therapeutic modality in this disease; on the other hand antihistamines have their place among many other therapeutic approaches in the treatment regimen of atopic eczema (11, 56, 68, 94).

New approaches include the application of mast

Table VII. Antihistamines, itch and atopic eczema

H1/H2 antagonists not superior to H1 or H2 alone (Foulds & Mackie, 1981)
Terfenadine, astemizol (1 dose) no effect upon endogenous, but upon peripherally induced itch (Krause & Shuster, 1983)
H1/H2 not superior to H1 but positive trend regarding itch (Frosch et al., 1984)
Histamine antagonism independent of sedation (Hägermark et al. 1985)
Tazyfylline no effect upon itch and scratch response in atopic eczema (Savin et al., 1986)
Tazyfylline dose-dependent effect upon peripherally induced itch (Ring et al., 1988)
Oxatomide effective in atopic eczema (placebo control) (Weinberg & Leaver, 1987)
Terfenadine, Ketotifen both effective equally (Tholen et al., 1987)
Subjective feeling of sedation independent of objective parameters (driving performance) (Ring & Bieber, 1987)
Dimetinden, astemizol equally effective (Kiehn & Rakoski, 1987)

cell blockers like oral cromoglycate, where we found some beneficial effect in an open clinical trial especially in patients with evidence for food allergy (45).

The modulation of fatty acid metabolism, either by giving gamma-linoleic acid (77, 93) or eicosapentaenoic acid (EPA) is under investigation. Our results with a double-blind controlled study with EPA in atopic eczema showed no effect compared to placebo (to be published).

Future studies in the field of mediator research may lead to new therapeutic approaches for the treatment of atopic eczema.

REFERENCES

1. Apold J, Aksnes L. Correlation between increased bronchial responsiveness to histamine and diminished plasma cyclic adenosine monophosphate response after epinephrine in asthmatic children. *J Allergy Clin Immunol* 1974; 59: 343-347.
2. Archer DB, Page CP, Juhlin L, Morley J, MacDonald M. Potentiation of inflammatory responses to prostaglandin E₂ in human skin by neutrophil chemoattractant phospholipids. *J Invest Dermatol* 1984, in press.
3. Archer CB, Morley J, MacDonald DM. Adrenoceptor function in atopic dermatitis: in vitro and in vivo observations. *Acta Derm Venereol*, in press.
4. Arrang J-M, Schwartz J-C, Schunack W. Stereospecificity of the histamine H₃-presynaptic autoreceptor. *Eur J Pharmacology* 1982; 117: 109-114.

5. Assem ESK, Attallah NA. Increased release of histamine by anti-IgE from leucocytes of asthmatic patients and possible heterogeneity. *Clin Allergy* 1981; 11: 367-373.
6. Atherton DJ. Allergy and atopic eczema I. *Clinical Exp Dermatol* 1981; 6: 191-203.
7. Austen KF. Biologic implications of the structural and functional characteristics of the chemical mediators of immediate-type hypersensitivity. New York: Academic Press, The Harvey Lectures, Series 1979; 73: 93-161.
8. Beaven MA, Jacobson S, Horakova Z. Modification of the enzymatic isotope assay of histamine and its application to measurement of histamine in measurement of histamine in tissues, serum and urine. *Clin Chem Acta* 1972; 37: 91-96.
9. Beer DJ, Rocklin RE. Histamine-induced suppressor-cell activity. *J Allergy Clin Immunol* 1984; 4: 439-452.
10. Bosse K, Hünecke P. Psychodynamik und Soziodynamik bei Hautkrankheiten. Göttingen, 1976.
11. Braun-Falco O, Ring J. Zur Therapie des atopischen Ekzems. *Hautarzt* 1984, in press.
12. Busse WW, Lee TP. Decreased adrenergic responses in lymphocytes and granulocytes in atopic eczema. *J Allergy Clin Immunol* 1976; 58: 586-596.
13. Busse WW, Lantis SDH. Impaired H₂ histamine granulocyte release in active atopic eczema. *Invest Dermatol* 1979; 73: 194-187.
14. Butler JM, Chan SC, Stevens SR, Hanifin JM. Increased leukocyte histamine release with elevated cyclic AMP-phosphodiesterase activity in atopic dermatitis. *J Allergy Clin Immunol* 1983; 71: 490-497.
15. Conroy MC, Adkinson NF, Lichtenstein LM. Measurement of IgE on human basophils: Relation to serum IgE and anti-IgE-induced histamine release. *J Immunol* 1977; 118: 1317-1324.
16. Czarnetzki BM. ECF, an eosinophil and neutrophil lipid chemotactic factor. *Behrin Inst Mitt* 1981; 58: 82-91.
17. Dorsch W, Ring J, Reimann HJ, Geiger R. Mediator studies in skin blister fluid from patients with dual skin reactions. *J Allergy Clin Immunol* 1982; 70: 236-244.
18. Dorsch W, Ring J, Melzer H. A selective inhibitor of thromboxane biosynthesis enhances immediate and reduces late cutaneous allergic reactions in man. *J Allergy Clin Immunol* 1983; 72: 168-174.
19. Dorsch W, Ring J, Riepel H. Effect of 15-Hydroxyeicosatetraenoic acid (15-HETE) on anti-immunoglobulin E- and calcium ionophore-induced histamine release from human leukocytes. *Int Arch Allergy Appl Immunol* 1984; 73: 274-279.
20. Dorsch W, Ring J, Weber PC, Strasser T. Detection of immunoreactive leukotrienes (LTC₄/D₄) in skin blister fluid after allergen testing in patients with late cutaneous reactions (LCR). 1984, in press.
21. Fantozzi R, Moroni F, Masini E, Blandina P, Mannaioni PF. Modulation of the spontaneous histamine release by adrenergic and cholinergic drugs. *Agents Actions* 1978; 8: 347-351.
22. Findlay SR, Lichtenstein LM. Basophil "releasability" in patients with intrinsic asthma. *J Allergy Clin Immunol* 1978; 61: 157.
23. Fiser PM, Buckley RH. Human IgE biosynthesis: in vitro studies with atopic and normal blood mononuclear cells and subpopulations. *J Immunol* 1979; 122: 1788.
24. Frosch PJ, Schwanitz HJ, Macher E. A double blind trial of H₁ and H₂ receptor antagonists in the treatment of atopic dermatitis. *Arch Dermatol Res* 1984; 276: 36-40.
25. Gallagher JS, Bernstein IL, Maccia CA, Splansky GL, Glueck HI. Cyclic platelet dysfunction in IgE-mediated allergy. *J Allergy Clin Immunol* 1978; 62: 229-235.
26. Gianetti A. High frequency of hereditary complement defects in association with atopic diseases. *Acta Dermatovener (Stockholm)* 1980; 92: 77.
27. Gillespie E, Valentine MD, Lichtenstein LM. Cyclic cAMP metabolism in asthma; studies with leukocytes and lymphocytes. *J Allergy Clin Immunol* 1974; 53: 27-33.
28. Grant JA, Dupree E, Thueson DO. Complement-mediated release of histamine from human basophils. *J Allergy Clin Immunol* 1977; 60: 306-311.
29. Grewe SR, Chan SC, Hanifin JM. Elevated leukocyte cyclic AMP-Phosphodiesterase in atopic disease. A possible mechanism for cAMP hyporesponsiveness. *J Allergy Clin Immunol* 1982; 70: 452-457.
30. Hanifin JM, Rogge JL, Bauman RH. Chemotaxis inhibition by plasma from patients with atopic dermatitis. *Acta Derm Venerol (Stockh)* 1980; 92: 52-56.
31. Hanifin JM, Rajka G. Diagnostic features of atopic dermatitis. *Acta Derm Venerol (Stockh)* 1980; 92: 44-47.
32. Hanifin JM. Clinical and basic aspects of atopic dermatitis. *Seminars in Derm* 1983; 2: 5-19.
33. Henderson WR, Smith L, Kaliner M. Alpha-adrenergic response in asthma. *Clin Res* 1978; 26: 227A.
34. Heskell N, Chan SC, Stevens SR, Hanifin JM. Leukocyte cAMP-Phosphodiesterase as a marker for atopy in neonates. *Clin Res* 1983; 31: 573.
35. Ishizaka K. Regulation of IgE response by IgE-binding factors. *Monogr Allergy* 1983; 18: 52-60.
36. Ishizaka T, Conrad DH. Binding characteristics of human IgE receptors and initial triggering events in human mast cells for histamine release. *Monogr Allergy* 1983; 18: 14-24.
37. Johnson HH, De Oreo GA, Lascheid WP, Mitchell F. Skin histamine levels in chronic atopic dermatitis. *Invest Dermatol* 1960; 34: 237-243.
38. Juhlin L. Localization and content of histamine in normal and diseased skin. *Acta Derm Venerol* 1967; 42: 218-226.
39. Juhlin L, Michaelsson G. Cutaneous vascular reactions to prostaglandins in healthy subjects and in patients with urticaria and atopic dermatitis. *Acta Derm Venerol* 1969; 49: 251-261.
40. Kaliner M. Human lung tissue and anaphylaxis. Evidence that cyclic nucleotides modulate the immunologic release of mediators through effects on microtubular assembly. *J Clin Invest* 1977; 60: 951-957.
41. Kaufmann HS, Frick OL, Fink D. Serum complement (β 1C) in young children with atopic dermatitis. *J Allergy* 1968; 42: 1-6.
42. Knauer A, Kagey-Sobotka A, Adkinson NF, Lichtenstein LM. Platelet augmentation of IgE-dependent histamine release from human basophils and mast cells. *Int Arch Allergy Appl Immunol* 1984; 74: 29-35.
43. Konzelmann M, Storck H. Wirkung von Cholinestern (Methacholin), Histamin, Kältereizen und psychischen Einflüssen auf Haut und Zirkulation von Neurodermati-

- tis-Patienten und Vergleichspersonen. *Hautarzt* 1965; 16: 304-310.
44. Korting GW. Zur Pathogenese des endogenen Ekzems. Stuttgart: Thieme, 1954.
 45. Kunz B, Vieluf D, Ring J. Efficacy of oral disodium cromoglycate (DSCG) in atopic eczema. XII. Int Congr Allergol Clin Immunol Abstr, Montroux, 1988.
 46. Lichtenstein LM, Margolis S. Histamine release in vitro: inhibition by catecholamines and methylxanthines. *Science* 1968; 161: 902-906.
 47. Lichtenstein LM, Schleimer RP, Peters SP, Kagey-Sobotka A, Adkinson NF, Adam GK, Schulman ES, MacGlashan DW. Studies with purified human basophils and mast cells. *Monogr Allergy* 1983; 18: 259-264.
 48. McDonald JR, Tan EM, Stevenson DD, Vaughan JH. Platelet aggregation in asthmatic and normal subjects. *J Allergy Clin Immunol* 1974; 54: 200-205.
 49. Makino S, Ikemori K, Kahima T, Fukuda T. Comparison of cyclic adenosine monophosphate response of lymphocytes in normal and asthmatic subjects to norepinephrine and salbutamol. *J Allergy Clin Immunol* 1977; 59: 348-352.
 50. Marone G, Kagey-Sobotka A, Lichtenstein LM. Effects of arachidonic acid and its metabolites on antigen-induced histamine release from human basophils in vitro. *Immunol* 1979; 123: 1669-1677.
 51. Marone G, Giugliano R, Lembo G, Ayala F. Human basophil releasability. II. Changes in basophil releasability in patients with atopic dermatitis. *J Invest Dermatol* 1986; 87: 19-23.
 52. Mihm MC Jr, Sotcr NA, Dvorak HF, Austen KR. The structure of normal skin and the morphology of atopic eczema. *J Invest Dermatol* 1976; 67: 305-308.
 53. Mikhail GR, Miller-Milenska A. Mast cell population in human skin. *J Invest Dermatol* 1976; 67: 305-308.
 54. Parker CW, Kennedy S, Eisen AZ. Leukocyte and lymphocyte cyclic AMP response in atopic eczema. *J Invest Dermatol* 1977; 68: 302-306.
 55. Pochet R, Delepesse G, De Maubeuge J. Characterization of β -adrenoreceptors on intact circulating lymphocytes from patients with atopic dermatitis. *Acta Derm Venereol (Stockh)* 1980; 92: 26-29.
 56. Rajka G. *Atopic dermatitis*. London: Saunders, 1975.
 57. Rajka G. Itch and IgE in atopic dermatitis. *Acta Derm Venereol (Stockh)* 1980; 92: 38-39.
 58. Rajka G. Recent therapeutic events: Cimetidine® and PUVA. *Acta Derm Venereol (Stockh)* 1980; 92: 117-118.
 59. Reed CE, Busse WW, Lee TP. Adrenergic mechanisms and the adenylyl cyclase system in atopic dermatitis. *J Invest Dermatol* 1976; 67: 333-338.
 60. Reimann JH, Meyer HJ, Wendt P. Stress and histamine. In: Ring J, Burg G, eds., *New trends in allergy*, pp. 50-56. Berlin: Springer, 1981.
 61. Ring J. Atopic dermatitis: a disease of general vasoactive mediator dysregulation. *Int Arch Allergy Appl Immunol* 1979; 59: 233-238.
 62. Ring J, Senter T, Cornell RC, Arroyave CH, Tan EM. Complement and immunoglobulin deposits in the skin of patients with atopic dermatitis. *Br J Dermatol* 1978; 99: 495-501.
 63. Ring J, Senter T, Cornell RC, Arroyave CM, Tan EM. Plasma complement and histamine changes in atopic dermatitis. *Br J Dermatol* 1978; 100: 521-526.
 64. Ring J. Zyklisches Adenosin-3-5-Monophosphat (cAMP) und Allergie. *Hautarzt* 1978; 29: 265.
 65. Ring J, Mathison DA, O'Connor R. In vitro cyclic nucleotide responsiveness of leukocytes and platelets in patients suffering from atopic dermatitis. *Int Arch Allergy Appl Immunol* 1981; 65: 1-7.
 66. Ring J. Plasma histamine concentrations in atopic eczema. *Clin Allergy* 1983; 13: 545-552.
 - 66a. Ring J, Dorsch W. Altered releasability of vasoactive mediator secreting cells in atopic eczema. *Acta Derm Venereol (Stockh)* 1985; 114: 9-23.
 67. Ring J, Palos E, Zimmermann F. Psychosomatische Aspekte der Eltern-Kind-Beziehung bei atopischem Ekzem: Psychodiagnostische Testverfahren bei Eltern und Kindern und Vergleich mit somatischen Befunden. *Hautarzt* 1985 (in press).
 68. Ring J. *Angewandte Allergologie*. 2. Auflage. MMW-Vieweg, München, 1988.
 69. Ring J, Sedlmeier F, von der Helm D, Mayr T, Walz U, Ibel H, Riepel H, Przybilla B, Reimann H-J, Dorsch W. Histamine and allergic diseases. In: Ring J, Burg G, eds. *New trends in allergy*. II, pp. 44-77. Berlin: Springer, 1988.
 70. Ring J, Przybilla B, Schwab U, Steger O. Klinisches Spektrum der Überempfindlichkeitsreaktionen gegen Sulfite. Fallbericht und Übersicht. *Allergologie* 1987; 3: 100-106.
 71. Ring J. Dermatologic diseases secondary to food allergy and pseudoallergy. *Food Allergy* 1988; 17: 271-.
 72. Rocklin RE, Greineder D. Histamine-induced suppressor factor (HSF): nature of stimulus and effect. *J Allergy Clin Immunol* 1978; 61: 144-150.
 73. Roszkowski W, Plaut M, Lichtenstein LM. Selective display of histamine receptors on lymphocytes. *Science* 1977; 195: 683-685.
 74. Ruzicka T, Glück S. Cutaneous levels and histamine releasability from the skin in atopic dermatitis and hyper-IgE-syndrome. *Arch Dermatol Res* 1983; 275: 41-44.
 75. Ruzicka T, Simmet T, Peskar BA, Braun-Falco O. Leukotrienes in skin of atopic dermatitis. *Lancet* 1984; 1: 222-223.
 76. Ruzicka T, Ring J. Enhanced releasability of prostaglandin E_2 and leukotrienes B_4 and C_4 from leukocytes of patients with atopic eczema. *Acta Derm Venereol (Stockh)* 1987; 67: 469-475.
 77. Schalin-Karrila M, Mattila L, Jansen CT, Uotila P. Evening primrose oil in the treatment of atopic eczema: effect on clinical status, plasma phospholipid fatty acids and circulating blood prostaglandins. *Br J Dermatol* 1987; 117: 11-19.
 78. Schmutzler W, Pobleto-Freundt G, Rauch K, Schoenfeld W. Response to immunological or cholinergic stimulation of isolated mast cells from man, guinea pig and rat. *Monogr Allergy* 14: 288-291.
 79. Schmutzler W, Delmich K, Eichelberg D, Glück S, Greven T, Jürgensen H, Riesener KP, Risse G, Pult P. The human adenoal mast cell. Susceptibility to different secretagogues and secretion inhibitors. *Int Arch Allergy Appl Immunol* 1985; 77: 177-178.

80. Schöpf E. Störung zellvermittelter Immunreaktionen bei Neurodermatitis atopica. Verminderte Spontanrosettenbildung von T-Lymphozyten. *Dermatologica* 1974; 149: 210–219.
81. Schwartz A, Sutton SL, Askenase PW, Gershon RK. Histamine inhibition of concanavalin A-induced suppressor T-cell activation. *Cell Immunol* 1981; 60: 425–439.
- 81 *a*. Schwartz A, Barbin G, Duchemin AM, Garbag M, Palacios JM, Quach TT, Rose C. Histamine receptors in the brain: characterization by studies and biochemical effects. In: Pepcu G, Kuhar MJ, Enna SJ, eds. *Receptors for neurotransmitters and peptides*, pp. 169–182. New York: Raven, 1980.
82. Simon RA, Stevenson DD, Arroyave CM, Tan EM. The relationship of plasma histamine to the activity of bronchial asthma. *J Allergy Clin Immunol* 1977; 60: 312–316.
83. Siraganian RP, Siraganian PA. Mechanism of action of concanavalin A on human basophils. *J Immunol* 1975; 114: 886.
84. Spornraft P, Przybilla B, Ring J. Histamine release after UVA-irradiation of human leukocytes in vitro, in press.
85. Strannegard IL, Lindholm L, Strannegard Ö. T lymphocytes in atopic children. *Int Arch Allergy Appl Immunol* 1976; 50: 684–692.
86. Sullivan TJ, Parker KL, Stenson W, Parker CW. Modulation of cyclic cAMP in purified rat mast cells. I. responses to pharmacologic, metabolic and physical stimuli. *J Immunol* 1975; 114: 1473–1479.
87. Szentivanyi A. The beta-adrenergic theory of the atopic abnormality in bronchial asthma. *J Allergy* 1968; 42: 203–232.
88. Szentivanyi A, Heim O, Schultz P, Szentivanyi J. Adrenoceptor bindings studies with 3-H (dihydroalprenolol) and 3-H (dihydroergocryptine) on membranes of lymphocytes from patients with atopic disease. *Acta Derm Venereol* 1980; 92: 19–22.
89. Thomas P, Ring J, Rieber P. In vitro IgE secretion in atopic eczema. *Acta Derm Venereol*, in press.
90. Venencie PY, Lebel B, Saurat JH. In: Ring J, Burg B, eds. *New trends in allergy*, pp. 224–230. Berlin: Springer, 1981.
91. Von der Helm D, Ring J, Dorsch W. Comparison of histamine release and prostaglandin E₂ production of human basophils in atopic and normal individuals. *Arch Dermatol Res* 1987; 279: 536–542.
92. Whitlock FA. *Psychophysiological aspects of skin disease*. London, Philadelphia, Toronto: Saunders, 1976.
93. Wright S, Burton JL. Oral-evening-primrose-seed oil improves atopic eczema. *Lancet* 1982; 1: 1120–1122.
94. Wüthrich B. *Zur Immunopathologie der Neurodermatitis constitutionalis*. Bern: Hans Huber, 1975.
95. Yamamoto K. Immunoglobulin, complement and fibronolytic enzyme system in atopic dermatitis. *Mod Probl Paediatr* 1975; 17: 130–135.