

Immunobiochemical Aspects of Atopic Dermatitis

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A long-term goal is to understand the pathomechanisms of atopic dermatitis. A major advance in the understanding of atopy was provided by observations from bone marrow transplantations which documented the transfer of the atopic diathesis by marrow cells (1). Conversely, the eczema of patients with Wiskott-Aldrich syndrome resolves after bone marrow transplantation (2). Thus, the immune and inflammatory cells that populate and infiltrate the skin in atopic dermatitis, or the nasal membranes in allergic rhinitis, or the bronchial mucosa in asthma, appear to be "vectors" that predispose the tissues to the hyperreactivity typical of the atopic diathesis.

A wide variety of factors have been reported to trigger flares of atopic dermatitis (3) (Table I). Of these, only stress and foods have been documented to cause flaring of dermatitis under controlled, experimental conditions (4, 5). Graham & Wolf reported increased skin temperature and decreased reactive hyperemia during experimental emotional stress interviews (4), a phenomenon frequently observed in the clinical setting. The mechanism for these reactions is not fully understood, but it seems likely that the initial event is the release of mediators from skin mast cells. Neuropeptides such as Substance P stimulate histamine release from skin mast cells and may link the central nervous system to cutaneous inflammatory cells (6, 7). Sampson has shown that double-blind food challenges cause itching and erythema accompanied by increased plasma histamine (5, 8) and later infiltration of eosinophils (9). Current concepts suggest that, following mast cell mediator release, infiltrates of basophils, eosinophils, neutrophils, and mononuclear leukocytes may interact to establish a continuing, subacute immune response (10). This may be a hybrid, with components of delayed hypersensitivity (11) and "late phase reactions" which could account for the chronic, indurated, inflammatory condition typical of AD.

In addition to evidence for abnormal inflammatory activity in atopic dermatitis, there are several lines of evidence indicating defects of chemotaxis and cellular immunity. However, our past studies of these abnormalities suggested they were secondary to the derma-

titis and normalized rapidly during clinical remissions (12, 13). Abnormalities of IgE are perhaps the most consistent immunological defect in atopic dermatitis. Serum IgE levels are elevated in approximately 80% of patients and correlate roughly with disease severity (14). Cultured mononuclear leukocytes, from patients with elevated serum IgE, produce excessive quantities of IgE during seven to ten day incubations and IgE production appears to be influenced by T cell factors but, as with inflammatory events, the mechanisms of this dysfunction remain to be clarified (15).

In addition to the *in vitro* IgE overproduction, another very consistent functional leukocyte abnormality in atopic dermatitis is the hyperreleasability of histamine by blood basophils (16-18). We have been interested in the cellular regulatory defects that allow for hyper-IgE production by B lymphocytes as well as the pathomechanism that allows for excessive basophil histamine release. A number of clinical clues, as well as certain *in vitro* findings, suggest this may relate to abnormal cyclic nucleotide metabolism in atopic dermatitis (19). The blunted cAMP response to catecholamines was initially interpreted as a beta-adrenergic receptor defect but we found no such abnormality in atopic leukocytes (20) and, along with other laboratories, we showed that this cyclic AMP defect was evident whether cells were stimulated with beta-agonists, prostaglandin (PG) or histamine (21).

These findings led us to the demonstration that reduced cAMP levels in stimulated mononuclear leukocytes (MNL) resulted from excessive hydrolysis by cAMP-phosphodiesterase (PDE) rather than inadequate cAMP production (22). This increased PDE activity was present consistently in MNL from pa-

Table I. *Confirmed and putative activators of atopic dermatitis*

Irritants	Immune complexes
Stress	Mites
Foods	Molds
Staphylococci	Yeasts
Viruses	Human dander

tients with active and inactive atopic dermatitis and also from patients with no dermatitis but only allergic respiratory disease. Non-atopic patients with widespread allergic contact dermatitis had normal levels of PDE activity (22).

Functional ramifications of increased leukocyte PDE activity were studied in two systems. The increased basophil histamine-releasability associated with atopic dermatitis showed a striking correlation to increased PDE activity and the abnormal histamine release was consistently reduced to normal levels by *in vitro* exposure of cells to the PDE inhibitor, RO 20-1724 (17). Likewise, the elevated IgE synthesis by cultured MNL from atopic dermatitis patients correlated with high PDE activity; exposure of the cells to Ro20-1724 for 1 hour, prior to the 10 day cultures, caused a consistent reduction in IgE synthesis (23).

Thus, excessive PDE hydrolysis of cAMP may have a functional role in IgE hyper-production and in basophil/mast cell hyper-releasability of mediators in atopic dermatitis. We have also been interested in abnormalities of other cell systems. We have focused especially on the blood monocyte, which has a particularly high level of PDE activity in atopic dermatitis (24), and is of major interest to our development of specific anti-PDE antibodies. Interestingly, our chromatofocusing studies have shown evidence of distinct PDE enzymes in atopic lymphocytes and monocytes, raising the possibility that different post-translational changes may be acting in the two cell types, or perhaps, in each of the many cell lines originating from bone marrow (25). Understanding these changes may potentially lead to development of a new therapeutic approach for atopy.

It is obvious from our studies that abnormally high PDE activity is present in atopic disease, in cells that are central to immune function. The resulting, inadequate cAMP levels would be expected to cause a permissive, functional hyper-reactivity which is certainly typical of the atopic diathesis. In therapeutic terms, these studies provide a focus for pharmacologic intervention and indeed, studies have shown the effectiveness of a topically applied PDE inhibitor (unpublished placebo-controlled trial). Additionally, our *in vitro* studies have shown that chronic, oral theophylline administration is ineffective for atopic dermatitis because of tachyphylaxis (26) and, perhaps, because of inadequate delivery of oral drug to the skin, since intravenous theophylline is rapidly effective at relieving the pruritus of atopic dermatitis (27).

A very important question relating to our research

is whether increased PDE activity reflects a basic biochemical genetic defect or whether underlying immunological events, possibly of allergic origin, cause elaboration of factors or differences in immune cellular differentiation which in turn generate a secondary rise in PDE activity. These questions are very basic but potentially have enormous medical and socio-occupational value, considering the substantial proportion of the population carrying the atopic diathesis.

REFERENCES

1. Saarinen UM. Transfer of latent atopy by bone marrow transplantation? A case report. *J Allergy Clin Immunol* 1984; 74: 196-200.
2. Saurat JH. Eczema in primary immune deficiencies. Clues to the pathogenesis of atopic dermatitis with special reference to the Wiskott Aldrich syndrome. *Dermatovenerol (Suppl)* 1985; 114: 125-128.
3. Hanifin JM. Atopic dermatitis. *J Allergy Clin Immunol* 1984; 73: 211-222.
4. Graham DT, Wolf S. The relation of eczema to attitude and vascular reactions of the human skin. *J Lab Clin Med* 1953; 42: 238.
5. Sampson HA. Role of immediate food hypersensitivity in the pathogenesis of atopic dermatitis. *J Allergy Clin Immunol* 1983; 71: 473-478.
6. Hägermark O, Hökfelt T, Pernow B. Flare and itch induced by substance P in human skin. *J Invest Dermatol* 1978; 71: 233-235.
7. Foreman JC. Peptides and histamine release. *J Allergy Clin Immunol* 1984; 74: 127-131.
8. Sampson HA, Jolie PL. Increased plasma histamine concentrations after food challenges in children with atopic dermatitis. *N Engl J Med* 1984; 311: 372-376.
9. Leiferman KM, Ackerman SJ, Sampson HA, et al. Dermal deposition of eosinophil-granule major basic protein in atopic dermatitis. *N Engl J Med* 1985; 313: 282-285.
10. Lemanske RF, Kaliner MA. Late phase allergic reactions. *Int J Dermatol* 1983; 22: 401-409.
11. Bruynzeel-Koomen C. IgE on Langerhans cells: New insights into the pathogenesis of atopic dermatitis. *Dermatologica* 1986; 172: 181-183.
12. Elliott ST, Hanifin JM. Delayed cutaneous hypersensitivity and lymphocyte transformation. Dissociation in atopic dermatitis. *Arch Dermatol* 1979; 115: 36-39.
13. Rogge JL, Hanifin JM. Immunodeficiencies in severe atopic dermatitis: Depressed chemotaxis and lymphocyte transformation. *Arch Dermatol* 1976; 112: 1391-1396.
14. Juhlin L, Johansson SGO, Bennich H. Immunoglobulin E in dermatoses: Levels in atopic dermatitis and urticaria. *Arch Dermatol* 1969; 100: 12-16.
15. Leung DYM, Geha RS. Immunoregulatory abnormalities in atopic dermatitis. *Clin Rev Allergy* 1986; 4: 67-86.
16. Lebel B, Venencie PY, Saurat JH, Soubrane C, Paupe J. Anti-IgE induced histamine release from basophils in

- children with atopic dermatitis. *Acta Derm Venereol* (Stockholm) 1980; 92: 57-59.
17. Butler JM, Chan SC, Stevens SR, Hanifin JM. Increased leukocyte histamine release with elevated cyclic AMP-phosphodiesterase activity in atopic dermatitis. *J Allergy Clin Immunol* 1983; 71: 490-497.
 18. Marone G, Giugliano R, Lembo G, Ayala F. Human basophil releasability. II. Changes in basophil releasability in patients with atopic dermatitis. *J Invest Dermatol* 1986; 87: 19-23.
 19. Hanifin JM. Pharmacophysiology of atopic dermatitis. *Clin Rev Allergy* 1986; 4: 43-65.
 20. Galant SP, Underwood S, Allred S, Hanifin JM. Beta-adrenergic receptor binding on polymorphonuclear leukocytes in atopic dermatitis. *J Invest Dermatol* 1979; 72: 330.
 21. Safko MJ, Chan SC, Cooper KD, Hanifin JM. Heterologous desensitization of leukocytes: A possible mechanism of beta adrenergic blockade in atopic dermatitis. *J Allergy Clin Immunol*. 1981; 68: 218-225.
 22. Grewe SR, Chan SC, Hanifin JM. Elevated leukocyte cyclic AMP-phosphodiesterase in atopic disease: A possible mechanism for cyclic AMP-agonist hyporesponsiveness. *J Allergy Clin Immunol* 1982; 70: 452-457.
 23. Cooper KD, Kang K, Chan SC, Hanifin JM. Phosphodiesterase inhibition by Ro 20-1724 reduces hyper-IgE synthesis by atopic dermatitis cells in vitro. *J Invest Dermatol* 1985; 84: 477-482.
 24. Chan SC, Grewe SR, Stevens SR, Hanifin JM. Functional desensitization due to stimulation of cyclic AMP-phosphodiesterase in human mononuclear leukocytes. *J Cyclic Nucleotide Research* 1982; 8: 211-224.
 25. Hanifin JM, Chan SC. Characterization of cAMP-phosphodiesterase as a possible laboratory marker of atopic dermatitis. *Drug Development Research*, 1988.
 26. Guistina TA, Chan SC, Thiel ML, Baker JW, Hanifin JM. Increased leukocyte sensitivity to phosphodiesterase inhibitors in atopic dermatitis: Tachyphylaxis after theophylline therapy. *J Allergy Clin Immunol* 1984; 74: 252-257.
 27. Epstein E. Theophylline ethylenediamine as an antipruritic agent. *Arch Dermatol* 1946; 53: 281-286.