



Characterization of the Oral and Gut Microbiota in Patients with Psoriatic Diseases: A Systematic Review

Tanja TODBERG^{1,2}, Hannah KAISER^{1,2}, Claus ZACHARIAE^{1,2}, Alexander EGEBERG^{1,2}, Anne-Sofie HALLING^{1,2} and Lone SKOV^{1,2}
¹Department of Dermatology and Allergy, Herlev and Gentofte Hospital, University of Copenhagen and ²Copenhagen Research Group for Inflammatory Skin (CORGIS), Hellerup, Denmark

Advances in technology have led to an increased number of studies investigating the microbiome in patients with psoriasis. This systematic review examined data regarding the oral and gut microbiota in patients with psoriasis and/or psoriatic arthritis and the effect of probiotics on the microbiota and severity of psoriasis. Of 1,643 studies, 23 were included (22 observational, 1 interventional). Studies examined the microbiota using culture or 16S rRNA gene sequencing analysis. All culture-based studies identified an increased presence of oral *Candida* in patients with psoriasis, whereas small variations in the oral microbiota were found in a 16S rRNA gene-based study. All 16S rRNA gene sequencing based studies agreed that the gut microbiota of patients with psoriatic disease differed from that of healthy controls, but the results were heterogeneous. Probiotics were associated with a significant improvement in the severity of psoriasis, but did not change microbiota. Overall, studies lacked relevant inclusion criteria and baseline information. In conclusion, the role of the microbiota in patients with psoriasis requires further investigation using more robust methods.

Key words: psoriasis; psoriatic arthritis; microbiota; immune system; probiotics.

Accepted Jul 14, 2021; Epub ahead of print Jul 15, 2021

Acta Derm Venereol 2021; 101: adv00512.

Corr: Tanja Todberg, Department of Dermatology and Allergy, Herlev and Gentofte Hospital, University of Copenhagen, Gentofte Hospitalsvej 15, DK-2900, Denmark. E-mail: tanja.todberg@regionh.dk

Psoriasis is a chronic inflammatory skin disease affecting 2–4% of the population (1). It is associated with several comorbidities, including psoriatic arthritis (PsA) and inflammatory bowel disease (IBD) (2). The pathogenesis of psoriasis is believed to involve an interplay between genetics, environmental markers, and the immune system, in which interleukin (IL)-23 and the Th17-derived cytokines IL-17 and IL-22 are considered to be the main drivers of inflammation (3).

The gastrointestinal system harbours trillions of microbial cells, with more than 9.9 million genes identified and, currently, the potential role of aberrant gut microbiota in inflammatory diseases is the focus of intense research, as the intestinal microbiota is known to have a critical function in the maturation and homeostasis of the immune system (4).

SIGNIFICANCE

Studies investigating the association between psoriasis and the microbiome have increased rapidly. This systematic review examined the role of the oral and gut microbiota and the effect of probiotics in patients with psoriasis and/or psoriatic arthritis. Twenty-three out of an initial total of 1,643 studies were included in the analysis. Of these, 22 studies were observational and 1 was interventional. The results showed increased presence of *Candida* in the oral cavity, and all studies examining the gut microbiota identified an altered microbiota in patients with psoriatic disease, but, overall, the results were heterogeneous. Probiotics were associated with a significant decrease in psoriasis severity, but the microbiota was unchanged. Further research is required into the role of the microbiome in patients with psoriasis.

Studies, including murine models of imiquimod induced-psoriasis, have shown that changes in gut bacteria are associated with increased severity of skin inflammation, supporting the linkage between a gut–skin axis (5). Likewise, in adult mice, dysbiosis caused by administration of antibiotics has been associated with a reduced Th17 response and decreased psoriasiform inflammation (6). This potential modulation of the gut microbiome may represent a new target for the treatment of psoriasis; however, a greater understanding of the functional potential of the gut microbiome is needed.

In recent years there has been increased focus on inflammation and psoriasis, leading to an emerging number of studies investigating the microbiome in patients with psoriasis. Methodological characterization of the microbiome has evolved rapidly; thus, culture-dependent methods are being replaced by sequencing analyses, such as 16S rRNA gene amplicon technique and shotgun metagenomic sequencing (7). Consequently, differences in study designs and methods make it difficult to interpret the results.

The focus of this systematic review was to summarize results, taking into account methodological variations in studies, regarding the gut microbiota in patients with psoriatic diseases (i.e. psoriasis or PsA) compared with healthy controls. Since the oral cavity is part of the gastrointestinal tract and, due to the association between psoriasis and periodontal diseases, oral microbiota studies were also included (8).

METHODS

This systematic review was established and performed in accordance with the Preferred Reporting Items for Systematic Review and Meta-Analyses (PRISMA) guidelines (9). A priori, a protocol was registered at PROSPERO (CRD42020168641).

Literature search

The databases PubMed, Embase, Cochrane Library and clinicaltrials.gov were searched for articles and trials until 11 March 2021, using the search term: “((psoriasis OR psoriatic OR psoriatic arthritis) AND ((microbiome OR microbiota OR microbial OR microflora OR prebiotics OR probiotics OR synbiotics) OR ((gut OR gastrointestinal OR gastro OR intestinal OR oral OR saliva OR dental) AND (diversity OR abundance OR composition OR balance)))”.

Study selection, outcome and quality assessment

Two authors (TT, HK) screened the titles and abstracts for eligible full-text studies. Eligibility criteria were observational studies that examined the association between patients with psoriasis and/or PsA and the oral and/or gut microbiome compared with healthy controls. Further interventional studies examining the effect of probiotics on the oral/and or gut microbiome in patients with psoriasis and/or PsA, were included. Animal studies and non-English studies, conference abstracts, case reports, and studies with no healthy control group or microbiome data were excluded. Newcastle Ottawa Scale (NOS) (score ≥ 7 indicated high-quality study) and Cochrane Collaboration's tool were used for quality assessment (10).

RESULTS

Study characteristics

A total of 1,643 studies were identified in the screening process. Of these, 105 full-texts were eligible for further review and 23 full-text articles were included in the analysis (**Fig. 1**).

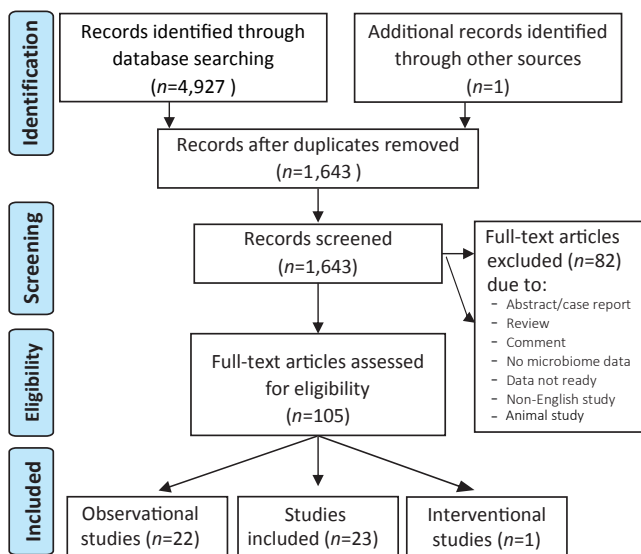


Fig. 1. Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) flow chart.

The studies included 4,379 participants, of whom 1,388 had psoriasis and 36 had PsA. All studies examined adults, except one study examining both children and adults (age range 10–82 years) (11) and, in most studies, the psoriatic population was age- and sex-matched with the control population (12–23). Nine studies included patients with plaque psoriasis only (11–13, 18, 20, 23–26), 6 studies included patients with mixed psoriasis types (15, 17, 21, 22, 27, 28) and 7 studies did not specify the type of psoriasis (14, 16, 19, 29–32). Fifteen studies examined patients with mild, moderate or severe psoriasis (11–13, 15–22, 24, 25, 27, 29), 2 studies examined patients with mild psoriasis (14, 32) and 4 studies reported no information on psoriasis severity (23, 28, 30, 31).

Quality assessment

Due to lack of information or unmatched controls, 16 studies were rated with a NOS score < 7 (11–13, 15, 17, 18, 22, 23, 25, 27–33), and 6 studies rated ≥ 7 (14, 16, 19–21, 24). The interventional study was rated with a low risk of bias (34).

Observational studies

Oral microbiota and psoriasis. Five studies investigated the oral microbiota in patients with psoriasis ($n=347$) compared with healthy controls ($n=669$) (11, 13, 24, 29, 30). Of these, 4 used culture-dependent methods and 1 used 16S rRNA gene sequencing analysis. Studies included both untreated patients and patients who used anti-psoriatic treatment, in 4 studies antibiotics were prohibited from 0–3 months before inclusion (11, 24, 29, 30), 2 studies had excluded patients with diabetes (11, 24), and in one study it was clinically verified that neither patients with psoriasis nor controls had periodontitis (30). Different methods were used for collection of oral material (**Tables I** and **SI¹**).

- **Culture-based studies.** All 4 culture-based studies showed a significantly higher prevalence and count of *Candida* species in patients with psoriasis, although the prevalence of *Candida* varied between studies (11, 13, 24, 29). One study found a positive correlation between *Candida* colonization and Psoriasis Area and Severity Index (PASI) score (24), while another study found that neither severity of psoriasis nor treatment modality were associated with *Candida* colonization (29). In all studies *Candida albicans* was the most common species isolated, both in patients with psoriasis and in healthy controls (**Tables I** and **SI¹**).
- **Sequencing-based study.** In the 16S rRNA gene sequencing based study the oral microbiota was examined in 27 patients with psoriasis compared with 52

¹<https://www.medicaljournals.se/acta/content/abstract/10.2340/00015555-3882>

Table I. Overview of observational studies characterizing the oral microbiota in patients with psoriasis and in healthy controls

Author (year) Location	Sample	Cases/ controls	Psoriasis type	Antipsoriatic treatment/other restrictions	Method	Main findings
Waldman* (2001) Israel (13)	Saliva	50/50	PQ	NS	Culture	The prevalence and count of <i>Candida</i> species were significantly higher in patients with psoriasis compared with healthy controls. No association between PASI score and quantity of <i>Candida</i> colonies was observed.
Bedair (2012) Jordan (29)	Swab, smear and oral-rinse	100/100	NS	All psoriasis treatment was allowed Antifungal/antibiotics were prohibited 2 months prior to study start	Culture	The prevalence and count of <i>Candida</i> species was significantly higher in patients with psoriasis compared with healthy controls (prevalence 69% vs 44%, $p < 0.001$). Prevalence of <i>Candida</i> carriers slightly higher among smokers than non-smokers in psoriasis (not significant), opposite with healthy controls (not significant). No significant difference in prevalence of <i>Candida</i> in treated/untreated patients with psoriasis.
Sarvtin (2014) Iran (11)	Swab	100/50	PQ	Corticosteroids and antibiotics were not allowed. No information on other treatment. Diabetes patients excluded	Culture	The prevalence and count of <i>Candida</i> species were significantly higher in patients with psoriasis compared with healthy controls (63% vs 24%, $p < 0.05$).
Lesan (2018) Iran (24)	Smear	70/70	PQ	All patients had never been treated with systemics. No antibiotics, antifungals or corticosteroids were allowed within 2 months prior to study start. Smokers and patients with systemic diseases excluded	Culture	The prevalence and count of <i>Candida</i> species were significantly higher in patients with psoriasis compared with healthy controls 20% vs 2.8%, $p = 0.002$. There was a significant positive association between PASI score and colony count ($p < 0.001$).
Belstrøm (2019) Denmark (30)	Swab and saliva	27/52	NS	Antibiotics were not allowed 3 months prior to study start	16s rRNA (V1–V3), 22 PCR cycles, Illumina Miseq Sequencing	α -diversity/relative abundance (Shannon's Diversity Index): No significant difference in predominant genera/species between the groups. β -diversity (PCoA): showed a random distribution within the groups (no clustering). Genera: <i>Streptococcus</i> , <i>Prevotella</i> , <i>Veillonella</i> and <i>Neisseria</i> were the most dominating in both groups. Species: <i>Prevotella melalogenica</i> and <i>Streptococcus salivarius</i> were the most dominating in both groups. 21 bacterial taxa at various levels differentiated between the groups.

*Article also included in Table II.

NS: not specified; PSO: psoriasis; PQ: plaque-type psoriasis; PCoA: principal coordinates analysis; PCA: principal component analysis.

healthy controls (30). No significance was found in α - or β -diversity, but a pair-wise comparison showed that 21 taxa differed between the groups (Tables I and SI¹).

Gut microbiota and psoriasis. Sixteen studies investigated the gut microbiota in patients with psoriasis ($n=812$) compared with healthy controls ($n=242$). Various criteria for use of concomitant treatment were allowed; in the majority of studies, antibiotics were prohibited from 2 weeks to 3 months before inclusion, but 4 studies gave no information on the use of antibiotics (13, 28, 32, 33). In most studies patients used various treatments for psoriasis, including topical corticosteroids, phototherapy and systemic anti-inflammatory treatment. However, in 4 studies, patients were systemic-naïve for 3 months prior to inclusion (12, 14, 16, 20). Regarding comorbidities, 4 studies included patients with diabetes (18–20, 31). Heterogeneous methods were used for collection of stool samples (Tables II and SI¹).

- **Culture-based studies.** Two studies, including 393 patients with psoriasis and 100 healthy controls, examined the presence of yeast in the gut using culture-based identification. In both studies, *Candida albicans* was the most abundant, with a significantly higher prevalence of *Candida* in patients with psoriasis (72% and 68%) compared with healthy controls (54% and 46%) (13, 28). One of the studies found that there was no correlation between PASI score and *Candida* colo-

nization (13). Concomitant treatment or comorbidities were not described in any of the studies.

- **Sequencing-based studies.** A total of 14 studies examined the gut microbiota in patients with psoriasis ($n=424$) compared with healthy controls ($n=387$) using sequencing-based analyses. All studies used 16S rRNA gene sequencing analysis, but with various hypervariable regions as target for PCR amplification and various sequencing platforms (Table I).

α -diversity

Various methods were used to assess α -diversity, and comparison between studies showed large variability (12, 14, 16, 17, 19–23, 25, 31). For patients with psoriasis; 2 studies reported a decreased diversity (16, 25), one found an increased diversity (12) and 6 studies found no significant differences between patients with psoriasis and healthy controls (17, 18, 21–23, 31). In addition, one of these studies, including 32 patients with psoriasis and 64 healthy controls, reported that sex, phototherapy, diet, alcohol, smoking, physical activity or the severity of psoriasis did not significantly affect the microbial profile (18). Notably, another study found no difference in α -diversity when investigating 55 patients with psoriasis compared with 27 healthy controls, but a lower richness in patients with more severe psoriasis was shown (20).

Table II. Overview of observational and interventional studies characterizing the gut microbiota in patients with psoriasis and/or psoriatic arthritis and in healthy controls

Author (year) Location	Sample	Cases/ Controls	Disease/ Psoriasis type	Antipsoriatic treatment/ other restrictions	Method	Main findings
Buslau (1997) Germany (28)	Stool	343/50	PQ	NS	Culturing	Prevalence and count of <i>Candida</i> species was higher in patients with psoriasis compared with healthy controls (68% vs 54%, <i>p</i> : NS).
Smith (1997) Scotland (33)	Stool	5/36*	PsA	NS	Culturing	The most abundant bacteria isolated from stool was <i>E. coli</i> for patients with PsA. The most abundant bacteria isolated from stool were <i>E. coli</i> , <i>Enterococci</i> , <i>Klebsiella oxytoca</i> and <i>Klebsiella pneumoniae</i> for healthy controls.
Waldman (2001) Israel (13)	Stool	50/50	PQ	NS	Culturing	Prevalence and count of <i>Candida</i> species were significantly higher in patients with psoriasis compared with healthy controls (72% vs 46%, <i>p</i> < 0.01). No association between PASI score and quantity of <i>Candida</i> colonies in stool was observed.
Codoner (2014) Spain (12)	Stool	52/52	PQ	Systemic psoriasis treatment was not allowed 3 months prior to study start Systemic antibiotics were not allowed 2 weeks prior to study start	16sRNA (V3-V4) Illumina Miseq	α -diversity (Shannon's Diversity Index): Significantly higher diversity in patients with psoriasis compared with healthy controls. β -diversity (PCA): Differences were observed between patients with psoriasis compared with healthy controls, but some healthy controls clustered in the psoriasis group. Phylum (PSO): \downarrow Bacteroidetes Genera (PSO): \uparrow <i>Faecalibacterium</i> , \downarrow <i>Bacteroides</i> \uparrow <i>Akkermansia</i> , \uparrow <i>Ruminococcus</i>
Scher (2015) USA (NY) (14)	Stool	15/16/17*	PQ PsA	Systemic antibiotics were not allowed 3 months prior to study start Extreme diet Patients were DMARD and biologic-naive IBD patients excluded	16s rRNA (V1-V2) 454 pyrosequencing	α -diversity (Shannon's Diversity Index): Significantly lower diversity in patients with psoriasis compared with healthy controls. β -diversity (PCA): Significantly separated groups. Phylum (PSO, PsA): \downarrow Bacteroidetes, \uparrow Firmicutes. Genera (PSO): \downarrow <i>Coprobacillus</i> (PSO), \downarrow Parabacteroides. Genera (PsA): \downarrow <i>Akkermansia</i> (PSA), \downarrow <i>Ruminococcus</i> (PSA), and \downarrow <i>Pseudobutyrvibrio</i> (PSA). Species (PSO, PsA): \downarrow <i>Coprococcus</i> species.
Eppinga (2016) Holland (27)	Stool	29/33	Mixed	Antibiotics were not allowed 8 weeks prior to study start	1 6S rRNA qPCR, 40 PCR cycles	Species (PSO): \downarrow <i>F. prausnitzii</i> , \uparrow <i>E. coli</i>
Eppinga (2017) Holland (15)	Stool	30/32/28	Mixed	Antibiotics were not allowed 8 weeks prior to study start Patients with psoriasis treated with dimethylfumarate (DMF, <i>n</i> = 28)	16S rRNA qPCR, 40 PCR cycles	Species (PSO, untreated): \downarrow <i>S. cerevisiae</i> compared with healthy controls and patients with psoriasis (DMF-treated)
Tan (2018) China (25)	Stool	14/14	PQ	Systemic anti-inflammatory treatment was not allowed	16s rRNA (V4), 30 PCR cycles, Illumina Miseq	α -diversity: (Chao1/ACE): No significant difference, but psoriasis group showed a slight decreased diversity. 70% OTUs were shared, 118 (healthy) and 135 (controls) OTUs were individual to the groups. β -diversity: Slightly separated groups. Phylum (PSO): \downarrow <i>Verrucomicrobia</i> , \downarrow <i>Tenericutes</i> Class (PSO): \downarrow <i>Mollicutes</i> , \downarrow <i>Verrucomicrobiae</i> Order (PSO): \downarrow <i>Verrucomicrobiales</i> , \downarrow RF39 Family (PSO): \uparrow <i>Bacteridaceae</i> , \uparrow <i>Enterococcaceae</i> Genus (PSO): \downarrow <i>Akkermansia</i> , \uparrow <i>Enterococcus</i> and \uparrow <i>Bacteroides</i> Species (PSO): \downarrow <i>Akkermansia muciniphila</i> , \uparrow <i>Clostridium citroniae</i>
Hidalgo-Cantabrana (2019) Spain (16)	Stool	19/20	NS	Systemic psoriasis treatment and antibiotics were not allowed 3 months prior to study start	16s rRNA (V2-V3), Ion 16S Metagenomics Kit	α -diversity (Chao1, Shannon's Diversity Index): Significant lower diversity in patients with psoriasis compared with healthy controls. β -diversity (PCoA): Significantly separated groups. Phylum (PSO): \downarrow Bacteroidetes, \downarrow Proteobacteria, \uparrow Actinobacteria, \uparrow Firmicutes Family (PSO): \uparrow (<i>Bifidobacteriaceae</i> , <i>Coriobacteriaceae</i> , <i>Lachnospiraceae</i> , <i>Clostridiales</i> FamilyXIII, <i>Eggerthellaceae</i> , <i>Peptostreptococcaceae</i> , <i>Ruminococcaceae</i> and <i>Erysipelotricaceae</i>), \downarrow (<i>Bacteroidaceae</i> , <i>Barnesiellaceae</i> , <i>Prevotellaceae</i> , <i>Tannerellaceae</i> , <i>Burkholderiaceae</i> , <i>Rikenellaceae</i> , <i>Lactobacillaceae</i> , <i>Streptococcaceae</i> , <i>Desulfovibrionaceae</i> , <i>Veillonellaceae</i> , <i>Marinifilaceae</i> , <i>Victivallaceae</i> and <i>Pasteurellaceae</i>). Genus (PSO): \uparrow <i>Bifidobacterium</i> , \uparrow <i>Blautia</i> , \uparrow <i>Collinsella</i> , \uparrow <i>Slackia</i> , \downarrow <i>Bacteroides</i> , \downarrow <i>Parabacteroides</i> , \downarrow <i>Barnesiella</i> , \downarrow <i>Alistipes</i> , \downarrow <i>Paraprevotella</i> .
Shapiro (2019) Israel (31)	Stool	24/22	NS	Systemic antibiotics were not allowed 3 months prior to study start	16s rRNA (V4) Illumina Miseq	α -diversity (Shannon's Diversity Index): No difference. β -diversity: Significantly separated groups. Phylum (PSO): \uparrow Firmicutes, \uparrow Actinobacteria \downarrow Bacteroidetes, \downarrow Proteobacteria (no change after correction for age/sex/BMI). Genus (PSO): \uparrow <i>Blautia</i> , \uparrow <i>Faecalibacterium</i> \downarrow <i>Prevotella</i> . Species (PSO): \uparrow <i>Ruminococcus gnavus</i> , \uparrow <i>Dorea formicigenerans</i> , \uparrow <i>Collinsella aerofaciens</i> \downarrow <i>Prevotella copri</i> .

Table II. (contd.)

Author (year) Location	Sample	Cases/ Controls	Disease/ Psoriasis type	Antipsoriatic treatment/ other restrictions	Method	Main findings
Chen (2019) Taiwan (18)	Stool	32/64	PQ	All psoriasis treatment was allowed Antibiotics and PPI were not allowed 1 months prior to study start	16s rRNA (V3-V4) Illumina Miseq	<p>α-diversity (Chao1, Shannon's Diversity Index): No difference between groups.</p> <p>β-diversity: Significant difference between treated, untreated and healthy controls.</p> <p>Significant difference between patients with psoriasis and healthy controls in group with BMI < 25 kg/m², not among BMI ≥ 25 kg/m². At OTU level significant difference between those using biologics vs biologic-naïve.</p> <p>Phylum level (PSO + healthy controls): dominated by: Bacteroidetes, Firmicutes Proteobacteriae, Bacteroidetes, Firmicutes.</p> <p>Phylum: ↓ Bacteroidetes, ↑ Firmicutes.</p> <p>Family level (PSO + healthy controls): dominated by <i>Bacteroidaceae</i>, <i>Prevotellaceae</i>, <i>Ruminococcaceae</i>, <i>Veillonellaceae</i> and <i>Lacnospiraceae</i>.</p> <p>(PSO): ↓ <i>Prevotellaceae</i>, ↓ <i>Ruminococcaceae</i>, ↑ <i>Veillonellaceae</i> and ↑ <i>Lacnospiraceae</i></p> <p>Genus (PSO): ↑ <i>Ruminococcus</i>, ↑ <i>Megasphaera</i>, ↑ <i>Dialister</i> ↓ <i>Sutterella</i>, ↓ <i>Paraprevotella</i>.</p> <p>Covariates: Sex, PASI score, phototherapy, arthritis, diet, alcohol, smoking did not affect abundance profile among group of psoriasis and healthy controls.</p>
Yeh (2019) Taiwan (19)	Stool	24/12/10	NS	Antibiotics and systemic corticosteroids/anti-psoriatic treatment were not allowed 1 month prior to study start Patients with psoriasis treated with secukinumab (n = 24) Patients with psoriasis treated with ustekinumab (n = 10)	16s rRNA (V3-V4) Illumina Miseq	<p>α-diversity: No significant difference between treated groups.</p> <p>β-diversity (PCoA): Significant alterations of microbiome at 6 months in secukinumab group, but not in ustekinumab group.</p>
Manasson (2020) USA (NY) (32)	Stool	15/15*	PsA	Biologic-naïve or brief exposure > 1 year previously.	16s rRNA (V4) Illumina Miseq/ Fungal: ITS1 region	<p>Order (PsA): ↑ <i>Clostridiales</i>, ↑ <i>Erysipelotrichiales</i> ↓ <i>Bacteroidales</i> (no further comparisons).</p>
Dei-Cas (2020) Argentina (20)	Stool	55/27	PQ	Systemic psoriasis treatment, phototherapy and antibiotics were not allowed 3 months prior to study start	16s rRNA (V3-V4) Illumina Miseq	<p>α-diversity (Chao1 index): no significant difference between patients with psoriasis and healthy controls.</p> <p>β-diversity: Significant (weighted Unifrac analyses)/not significant (non-weighted Unifrac analyses).</p> <p>Phylum: No differences between subtypes, but difference in abundance.</p> <p>↓ Bacteroidetes (47.1% PSO vs 59.9% healthy controls), ↑ Firmicutes, (44.6% PSO vs 33% healthy controls) ↑ Proteobacteriae (5.4% PSO vs 4.2% healthy controls) ↑ Actinobacteriae (0.8% PSO vs 0.8% healthy controls)</p> <p>Genus (PSO): ↑ <i>Blautia</i>, ↑ <i>Faecalibacterium</i> ↓ <i>Paraprevotella</i>, ↓ <i>Bacteroides</i>.</p> <p>No significant changes in gut microbiota associated with change in age, weight and BMI.</p> <p>Mild vs moderate-to-severe psoriasis (PASI ≥ 10), patients with moderate to severe psoriasis had lower biodiversity than patients with mild psoriasis.</p>
Yegorov (2020) Kazakstan (21)	Stool	14/7	Mixed	Antibiotics were not allowed 3 months prior to study start	16s rRNA (V1-V3) Illumina	<p>α-diversity (Shannon's Diversity Index): No difference between groups</p> <p>Phylum: No differences in Firmicutes/Bacteroidetes ratio</p> <p>Family (PSO): ↓ <i>Lacnospiraceae</i> ↑ <i>Ruminococcaceae</i> Genus (PSO): ↑ <i>Faecalibacterium</i> ↓ <i>Oscillibacter</i></p>
Zhang (2021) China (22)	Stool	30/30	Mixed	Antibiotics were not allowed 1 month prior to study start	16sRNA (V3-V4) HiSeq platform	<p>α-diversity (Shannon's Diversity Index): No difference between groups</p> <p>β-diversity (non-weighted Unifrac analyses): Differences were observed between patients with psoriasis compared with healthy controls,</p> <p>Family (PSO): ↑ <i>Veillonellaceae</i>, ↑ <i>Ruminococcaceae</i> Genus (PSO): ↑ <i>Faecalibacterium</i> ↑ <i>Megamonas</i></p>
Wang (2021) China (23)	Stool	20/20	PQ	Psoriasis treatment and antibiotics not allowed 1 month prior to study start	16sRNA (V4) Ion5s platform	<p>α-diversity: (Chao/ACE, Shannons's/Simpsons diversity): No significant difference, but psoriasis group showed slight decreased diversity.</p> <p>β-diversity (PCoA): Significant separation of communities between patients with psoriasis and controls</p> <p>Genus (PSO): ↑ <i>Megamonas</i> ↓ <i>Rombutsia</i></p>

NS: not specified, PSO: psoriasis, PQ: plaque-type psoriasis; PCoA: principal coordinates analysis, PCA: principal component analysis.

*Patients with psoriatic arthritis (PsA).

β -diversity

In all 10 studies that assessed β -diversity, a difference was observed between patients with psoriasis and healthy controls (12, 14, 16–20, 22, 23, 25, 31). Interestingly, one study (18) found that the difference was present only when comparing patients with psoriasis ($n=19$) and healthy controls ($n=36$) with body mass index (BMI) <25 kg/m², but not among subjects with BMI ≥ 25 kg/m².

Differences in relative abundance

Studies showed large variability when evaluating relative abundance and focused on various taxonomy levels. At phyla level, 5 studies described Bacteroidetes, Firmicutes, Proteobacteria and Actinobacteria as the most dominating bacteria in both patients with psoriasis and in healthy controls, with Bacteroidetes and Firmicutes as the most abundant (16–18, 20, 31).

In patients with psoriasis, 6 studies found Bacteroidetes to be decreased, and Firmicutes and Actinobacteria to be increased (12, 14, 16, 18, 20, 31); however, one study found the opposite, with an increased level of Bacteroidetes and decreased levels of Firmicutes and Actinobacteria in patients with psoriasis ($n=35$) compared with healthy controls ($n=27$) (17). At the family level, 2 studies described that patients with psoriasis had less *Prevotellaceae* than healthy controls (16, 18). At the genus level, *Akkermansia* was increased in patients with psoriasis in one study, whereas a low abundance of *Akkermansia* in patients with psoriasis was found in 2 studies (12, 14, 25).

Gut microbiota and psoriatic arthritis. Three studies examined the gut microbiota in patients with PsA ($n=36$) compared with healthy controls ($n=68$) (14, 32, 33). In 2 studies, all patients also had psoriasis (14, 32). A culture-based study found *E. coli* isolated as the only species in a small group of patients with PsA ($n=5$), compared with the control group ($n=36$) where *E. coli*, *Enterococcus*, *Klebsiella oxytoca* and *Klebsiella pneumoniae* were present (33). Two studies were based on 16S rRNA gene analysis (14, 32), with one study presenting a lower abundance of *Akkermansia* in patients with PsA ($n=16$) compared with healthy controls ($n=17$) (14).

Interventional studies

Only one study examined the effect of probiotics on the gut microbiome and the severity of psoriasis. In this 12-week randomized controlled trial, 90 patients with psoriasis were randomized to daily doses of either probiotic capsules or placebo (34). Topical treatment was allowed during the study. At the end of the trial the authors found no significant difference in the microbiota between the probiotic and the placebo groups, but the results showed a significantly clinical effect on severity of psoriasis; 66.7% of subjects in the probiotic group achieved at least PASI75 compared with 41.9% of subjects in the placebo group ($p=0.03$) (Tables III and SII¹).

DISCUSSION

Main findings

Overall, this review demonstrates that patients with psoriasis seem to have some variations in their microbiota compared with healthy controls, but, in general, the results were based on small sample sizes and, across studies, there were wide variations in inclusion criteria and methods used.

Across studies using the culture-based method the prevalence of *Candida* in the oral cavity and gut was found to be higher in patients with psoriatic diseases, whereas the single study examining the oral microbiota using 16S rRNA gene sequencing analysis found no difference in α - or β -diversity, although some taxa differed between groups.

Across studies that examined the gut microbiota using 16S rRNA gene sequencing analysis, no core microbiota of patients with psoriatic disease was identified, although all the studies agreed that patients with psoriatic disease had a distinct microbiota compared with healthy controls. Except for β -diversity, where clustering was identified between patients with psoriatic disease and healthy controls, no consensus was found for α -diversity and lower taxonomic profiles varied between studies.

Interestingly, the interventional study found no significant change in microbial composition after administration of probiotics, but a considerable clinical improvement in psoriasis was seen.

Table III. Overview of interventional study characterizing the gut microbiome in patients with psoriasis after treatment with probiotics vs placebo

Author (year) Location	Sample	Cases/ Controls	Psoriasis type	Baseline PASI mean \pm SD	Antipsoriatic treatment/ other restrictions	Method	Main findings
Navarro-Lopez (2019) Spain (34)	Stool	46/44	PQ	11.7 \pm 5.1 11.5 \pm 4.2	Systemic psoriasis treatment was not allowed 3 months prior to study start Systemic antibiotics were not allowed 2 weeks prior to study start	16S rRNA (V3–V4) Illumina Miseq	α -diversity (Shannon's Diversity Index): No significance in Shannon's Diversity Index between groups. No statistically significant change microbial profile after treatment, but disappearance of the genera Micromonospora, Rhodococcus, increase in Collinsella and Lactobacillus in probiotic group. PASI75 week 12; Probiotic group: 66.7% vs placebo group: 41.9%.

NS: not specified; PSO: psoriasis; PQ: plaque-type psoriasis; PCoA: principal coordinates analysis; PCA: principal component analysis.

Interpretation

Several factors might contribute to the increased presence of *Candida* in the oral microbiota in patients with psoriasis. Patients with psoriasis use various immunosuppressive drugs, are more often smokers, and diabetes is more common among this patient group (35). In fact, in former studies, smoking and obesity have been associated with periodontitis (36, 37), which is characterized by an altered oral microbiota (38); hence conditions other than psoriasis may impact the oral microbiology. Of note, only one study verified that patients with psoriasis did not have periodontitis prior to inclusion (30).

In addition, food substances, such as simple carbohydrates, are known to affect the microbial environment, and may increase the risk of *Candida* colonization; however, studies gave no information about variations in diet between patients with psoriasis and healthy controls (39). Notably, 1 included study observed a distinct oral bacterial environment in patients with psoriasis compared with healthy controls and, as it is well known that approximately 1,500 bacterial species inhabit the oral cavity, with each person harbouring approximately 200 species, it cannot be excluded that patients with psoriasis have a distinct bacterial environment that is more susceptible to *Candida* species (30), although the results may be confounded by various factors.

All sequencing-based studies assessing the gut microbiota found heterogeneous results, although, at phyla level, 5 studies agreed that Bacteroidetes, Firmicutes, Proteobacteria and Actinobacteria were the most abundant, both in patients with psoriasis and in healthy controls (16–18, 20, 31). In support of this, previous studies have detected these phyla as the most dominating independent of health status (40, 41). It is widely discussed whether the dominance of bacterial groups may impact on health, with some reporting wide variation in dominance of, for example, the genus *Bacteroides*, in a large study of healthy individuals (42), indicating that a low or high ratio of specific families, genera or species may not indicate that a person is unhealthy or “dysbiotic”. In contrast, others have suggested that the dominance of specific phyla may be important, as previous studies have linked a high Firmicutes/Bacteroidetes ratio with obesity, while a decrease in the Firmicutes/Bacteroidetes ratio has been observed in patients undergoing weight loss (43). Notably, 6 studies agreed that Firmicutes were more abundant compared with Bacteroidetes in patients with psoriasis, one study found a lower Firmicutes/Bacteroidetes ratio, and one study found no significant difference in Firmicutes/Bacteroidetes ratio between patients with psoriasis and healthy controls (21).

It is well described that obesity is associated with an aberrant microbial profile; therefore BMI is an important factor to consider in the analyses of microbiota

data (44). In support of this, one study identified a difference between normal-weight patients with psoriasis and normal-weight healthy controls, but not between overweight groups; thus data from normal-weight subjects may represent a more “true” microbiota profile, when not confounded by obesity (18). Overall, patients in the studies had a mean BMI ≥ 25 kg/m² (15, 18, 20, 27, 31, 32). This is in line with the literature, as it is well known that patients with psoriasis are more often obese compared with the background population (45).

In one study the genus *Akkermansia* was found to be increased in patient with psoriasis (12), whereas 2 studies found that *Akkermansia* was decreased in patients with psoriasis, and decreased even more in patients with both psoriasis and PsA (14, 25). *Akkermansia* is a commensal in the large intestines, representing 1–4% of the microbiota in healthy adults, and has been associated with dual functions. In murine studies *Akkermansia* has been linked with mucus degradation (46, 47), which may, in turn, facilitate increased permeability of the intestinal wall. In theory, this could potentially contribute to the systemic inflammation of psoriasis, as bacterial DNA with origin from the intestines has been observed in peripheral blood in patients with moderate-to-severe plaque psoriasis, but not in healthy controls (48). In contrast, adequate levels of *Akkermansia* species have been associated with beneficial regulation of mucus thickness, interaction with the immune system via Toll-like receptors and with the production of short-chained-fatty-acids (SCFA), which are produced by gut microbiota as metabolites of a fibre-rich diet (49–51). SCFA acetate, propionate and butyrate are known to have immune regulatory effect on regulatory T-cells (Treg) in immune-mediated diseases (52). Of note, most of the main producers of butyrate belong the families of *Ruminococcaceae* *Lachnospiraceae* (53). A recent study has shown that, in murine models of imiquimod-induced psoriasis and in isolated peripheral human blood from patients with psoriasis, butyrate stimulated the activity of Treg, leading to reduced expression of IL-17 and IL-6 and increased IL-10 expression (54). Thus, a fibre-rich diet is likely to decrease proinflammatory metabolites in the gut, leading to reduced IL-17 expression, and thereby decreased severity of psoriasis. Only one study assessed diet, and found that this did not affect the microbial profile (18). However, diet has been shown to affect the microbiota in several studies (4, 49, 55). Indeed, the genus *Prevotella* has been linked to a diet rich in plant-fibre and is more common in non-Westernized populations and, in 4 studies, the abundance of *Prevotella* was decreased in patients with psoriasis compared with the control populations (16, 18, 20, 31).

The role of probiotics in psoriasis

In the interventional study, no significant change in the microbiota was observed in the probiotic-treated group

compared with the placebo group, but when evaluating the clinical effect, a substantial reduction in the severity of psoriasis was seen in the treated group, with 66.7% of patients achieving PASI75 compared with 41.9% of patients in the placebo group.

The gut microbiota comprises approximately 10^{12} microbial cells, compared with probiotics comprising 10^9 colony-forming units; thus the microbiota outnumbers the probiotics extensively (56). Probiotic capsules are often consumed orally, and therefore need to survive the low pH in the stomach and to be established in a very diverse ecosystem with competition from the commensal microbiota. Although authors found a significant difference in the clinical presentation of psoriasis, a limitation was that topical corticosteroid use was not prohibited. Notably, numerous studies in patients with other indications have shown that there is little evidence to show that probiotics can change the gut microbiota (57, 58).

Heterogeneity of study design

The overall results were heterogeneous, although in most studies patients were age- and sex-matched. In general, studies lacked description of possible factors that may interfere with the results. Antibiotics that are known to affect the microbiota were prohibited for at least 3 months in only 5 studies (14, 16, 20, 30, 31). A recent paper has shown the microbiome to be affected by antibiotics for up to 6 months (59).

In addition, possible confounders that may affect the results include; non-BMI-matched populations, inclusion of participants with other immune-mediated disease and concomitant treatments, such as metformin and proton pump inhibitors, which are well-known modulators of the microbiota (60, 61). When including patients with psoriasis in systemic treatment for psoriasis, authors should be aware that it may not be the association between the gut microbiota and psoriasis that is investigated, but rather the effect of a drug on the microbiota and psoriasis (60). Likewise, the collection of samples, season, ethnicity and matching groups from the same district can play a role (59).

Heterogeneity of methodology

In addition, the heterogeneity of the results may be explained by the type of analyses carried out. Culturing-based analysis is able to identify both bacteria and fungi, depending on the choice of media; however, as anaerobic bacteria comprise a large proportion of the commensals in the distal colon, this type of analysis is associated with several limitations, with only 0.1–10% of gut commensals being able to be grown in culture (62), thus a large discrepancy between culture- and sequenced-based studies exists, making them difficult to compare.

The 16S rRNA gene analysis targets both aerobic and anaerobic bacteria; however, the analysis is limited by

typically not being able to sequence deeper than genus level, and by fungi not being identified. In addition, it is often based on operational taxonomic units (OTUs), in which a 97% similarity profile of base pairs is grouping OTUs together, which may be associated with errors (59).

In the 16S rRNA gene sequencing-based studies, choice of primers varied. Choosing the right primers is crucial when conducting a microbiome study, i.e. a study showed that the V1–V2 primer was unable to identify the most abundant bacteria in the vaginal tract (63). To study the microbiota V4–V5 is recommended prior to V3–V4, as, in most cases, it produces the most comparable results (64, 65).

None of the studies used shotgun metagenomic sequencing, which may be because this is a relatively new method and the expensive price. Shotgun metagenomic sequencing detects bacteria at the species level and, sometimes, even at strain level, depending on the sequencing depth, and information is given about the functional potential of the microbiota (59). The key to improving understanding of the microbiome may not be to identify which bacteria are present, which, to some extent, is provided by the 16S rRNA gene sequencing analysis, but rather to recognize the function of the bacteria.

Studies investigating other inflammatory diseases, such as atopic dermatitis, found that, although the same species are identified in both patients and in healthy controls, at strain level the bacteria can be different, highlighting the limitations of using 16S rRNA gene sequencing analysis (66). Another strength of using shotgun metagenomic sequencing is that fungi, virus and phages can sometimes be detected, which are all commensals with affinity to interfere with bacteria (67).

Study limitations

This systematic review has several limitations, including risk of publication bias, and the identified differences between patients with psoriasis and healthy controls may be explained by various confounders. The studies are based on culture or 16S rRNA gene sequencing analysis, and therefore the function of the microbiota is not described. The gut microbiota was assessed using stool samples; however, this analysis is limited as it represents only the end-products of the microbial composition. Furthermore, the data are based on observational single-point time profiles, and how to utilize these findings in development of treatment strategies and clinical practice remains unclear.

Conclusion

This review found a difference in the oral and gut microbiota between patients with psoriatic diseases and healthy controls. However, overall, studies lacked important information, as restrictions due to treatments and included patients with comorbidities known to affect the microbiota. Methodological heterogeneity was found between

studies, including variations in sampling and processing of the identification of microbiota profiles; thus, any conclusions must be considered with caution. To date, there is no consensus for conducting a microbiome study; however, some guidelines have been published (59). In general, a challenge in microbiome studies is that it is a new research field. To improve insights of the microbiota in patients with psoriatic diseases, the design of future studies would benefit from strict matching, more restrictions, and assessment of the microbial profile with combined data and more advanced methods.

ACKNOWLEDGEMENTS

This study was supported by Aage Bangs Foundation.

Disclosures and conflicts of interests: TT has been an investigator for Novartis, Abbvie, Dr Wolff and Almirall. AE has received research funding from Pfizer, Eli Lilly, Novartis, Bristol-Myers Squibb, AbbVie, Janssen Pharmaceuticals, the Danish National Psoriasis Foundation, the Simon Spies Foundation, and the Kgl Hofbundtmager Aage Bang Foundation, and honoraria as consultant and/or speaker from AbbVie, Almirall, Leo Pharma, Galápagos NV, Sun Pharmaceuticals, Samsung Bioepis Co., Ltd., Pfizer, Eli Lilly and Co., Novartis, Galderma, Dermavant, UCB, Mylan, Bristol-Myers Squibb, and Janssen Pharmaceuticals. CZ has been an advisor, investigator and speaker for Abbvie, Eli Lilly, Novartis, Sanofi, Leo Pharma, UCB, CSL and Almirall. ASHO has received honoraria as speaker for Leo Pharma and consultant for Coloplast A/S. LS has been an advisor, investigator and speaker for Abbvie, Eli Lilly, Novartis, Sanofi, Celgene, Leo Pharma, BMS, UCB and Almirall, outside the submitted work. LS reports non-financial support from Abbvie, Sanofi, Janssen and grants from Novartis, Janssen, BMS and Sanofi.

REFERENCES

- Parisi R, Iskandar IYK, Kontopantelis E, Augustin M, Griffiths CEM, Ashcroft DM. National, regional, and worldwide epidemiology of psoriasis: systematic analysis and modelling study. *BMJ* 2020; 369: m1590.
- Alinaghi F, Tekin HG, Burisch J, Wu JJ, Thyssen JP, Egeberg A. Global prevalence and bidirectional association between psoriasis and inflammatory bowel disease – a systematic review and meta-analysis. *J Crohn's Colitis* 2020; 14: 351–360.
- Griffiths CEM, Armstrong AW, Gudjonsson JE, Barker JNWN. Psoriasis. Vol. 397, *Lancet* 2021; 397: 1301–1315.
- Lynch S V., Pedersen O. The human intestinal microbiome in health and disease. *N Engl J Med* 2016; 375: 2369–2379.
- Zákostelská Z, Málková J, Klimešová K, Rossmann P, Hornová M, Novosádová I, et al. Intestinal microbiota promotes psoriasis-like skin inflammation by enhancing Th17 response. *PLoS One* 2016; 11: e0159539.
- Zanvit P, Konkel JE, Jiao X, Kasagi S, Zhang D, Wu R, et al. Antibiotics in neonatal life increase murine susceptibility to experimental psoriasis. *Nat Commun* 2015; 6: 2824.
- Mas-Lloret J, Obón-Santacana M, Ibáñez-Sanz G, Guinó E, Pato ML, Rodríguez-Moranta F, et al. Gut microbiome diversity detected by high-coverage 16S and shotgun sequencing of paired stool and colon sample. *Sci Data* 2020; 7: 92.
- Egeberg A, Mallbris L, Gislason G, Hansen PR, Mrowietz U. Risk of periodontitis in patients with psoriasis and psoriatic arthritis. *J Eur Acad Dermatol Venereol* 2017; 31: 288–293.
- Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gøtzsche PC, Ioannidis JPA, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate healthcare interventions: explanation and elaboration. *BMJ* 2009; 339: b2700.
- Higgins JPT, Altman DG, Gøtzsche PC, Jüni P, Moher D, Oxman AD, et al. The Cochrane Collaboration's tool for assessing risk of bias in randomised trials. *BMJ* 2011; 343: d5928.
- Taheri Sarvtin M, Shokohi T, Hajheydari Z, Yazdani J, Hedayati MT. Evaluation of candidal colonization and specific humoral responses against *Candida albicans* in patients with psoriasis. *Int J Dermatol* 2014; 53: e555–560.
- Codoñer FM, Ramírez-Bosca A, Climent E, Carrión-Gutiérrez M, Guerrero M, Pérez-Orquín JM, et al. Gut microbial composition in patients with psoriasis. *Sci Rep* 2018; 8: 3812.
- Waldman A, Gilhar A, Duek L, Berdicevsky I. Incidence of *Candida* in psoriasis—a study on the fungal flora of psoriatic patients. *Mycoses* 2001; 44: 77–81.
- Scher JU, Ubeda C, Artacho A, Attur M, Isaac S, Reddy SM, et al. Decreased bacterial diversity characterizes the altered gut microbiota in patients with psoriatic arthritis, resembling dysbiosis in inflammatory bowel disease. *Arthritis Rheumatol* 2015; 67: 128–139.
- Eppinga H, Thio HB, Schreurs MWJ, Blakaj B, Tahitu RI, Konstantinov SR, et al. Depletion of *Saccharomyces cerevisiae* in psoriasis patients, restored by Dimethylfumarate therapy (DMF). *PLoS One* 2017; 12: e0176955.
- Hidalgo-Cantabrana C, Gomez J, Delgado S, Requena-Lopez S, Queiro-Silva R, Margolles A, et al. Gut microbiota dysbiosis in a cohort of patients with psoriasis. *Br J Dermatol* 2019; 181: 1287–1295.
- Huang L, Gao R, Yu N, Zhu Y, Ding Y, Qin H. Dysbiosis of gut microbiota was closely associated with psoriasis. *Sci China Life Sci* 2019; 62: 807–815.
- Chen Y-J, Ho HJ, Tseng C-H, Lai Z-L, Shieh J-J, Wu C-Y. Intestinal microbiota profiling and predicted metabolic dysregulation in psoriasis patients. *Exp Dermatol* 2018; 27: 1336–1343.
- Yeh N-L, Hsu C-Y, Tsai T-F, Chiu H-Y. Gut microbiome in psoriasis is perturbed differently during secukinumab and ustekinumab therapy and associated with response to treatment. *Clin Drug Investig* 2019; 39: 1195–203.
- Dei-Cas I, Giliberto F, Luce L, Dopazo H, Penas-Steinhardt A. Metagenomic analysis of gut microbiota in non-treated plaque psoriasis patients stratified by disease severity: development of a new Psoriasis-Microbiome Index. *Sci Rep* 2020; 10: 12754.
- Yegorov S, Babenko D, Kozhakhmetov S, Akhmaltdinova L, Kadyrova I, Nurgozhina A, et al. Psoriasis is associated with elevated gut IL-1 α and intestinal microbiome alterations. *Front Immunol* 2020; 11: 571319.
- Zhang X, Shi L, Sun T, Guo K, Geng S. Dysbiosis of gut microbiota and its correlation with dysregulation of cytokines in psoriasis patients. *BMC Microbiol* 2021; 21: 78.
- Wang X, Zhai W, Ma J, Xu S, Liu M, Zhang X, et al. Substantial alterations of the intestinal microbiota in psoriasis patients of China. *Exp Dermatol* 2021 Feb 3. [Online ahead of print].
- Lesan S, Toosi R, Aliakbarzadeh R, Daneshpazhooh M, Mahmoudi L, Tavakolpour S, et al. Oral *Candida* colonization and plaque type psoriasis: is there any relationship? *J Investig Clin Dent* 2018; 9: e12335.
- Tan L, Zhao S, Zhu W, Wu L, Li J, Shen M, et al. The Akkermansia muciniphila is a gut microbiota signature in psoriasis. *Exp Dermatol* 2018; 27: 144–149.
- Navarro-Lopez V, Martinez-Andres A, Ramirez-Bosca A, Ruzafa-Costas B, Nunez-Delegido E, Carrion-Gutierrez MA, et al. Efficacy and safety of oral administration of a mixture of probiotic strains in patients with psoriasis: a randomized controlled clinical trial. *Acta Derm Venereol* 2019; 99: 1078–1084.
- Eppinga H, Sperna Weiland CJ, Thio HB, van der Woude CJ, Nijsten TEC, Peppelenbosch MP, et al. Similar depletion of protective *Faecalibacterium prausnitzii* in psoriasis and inflammatory bowel disease, but not in hidradenitis suppurativa. *J Crohns Colitis* 2016; 10: 1067–1075.
- Buslau M, Menzel I, Holzmann H. Fungal flora of human faeces in psoriasis and atopic dermatitis. *Mycoses* 1990; 33: 90–94.
- Bedair AA, Darwazeh AMG, Al-Aboosi MM. Oral *Candida*

- colonization and candidiasis in patients with psoriasis. *Oral Surg Oral Med Oral Pathol Oral Radiol* 2012; 114: 610–615.
30. Belstrom D, Eiberg JM, Enevold C, Grande MA, Jensen CAJ, Skov L, et al. Salivary microbiota and inflammation-related proteins in patients with psoriasis. *Oral Dis* 2020; 26: 677–687.
 31. Shapiro J, Cohen NA, Shalev V, Uzan A, Koren O, Maharshak N, et al. Psoriatic patients have a distinct structural and functional fecal microbiota compared with controls. *J Dermatol* 2019; 46: 595–603.
 32. Manasson J, Wallach DS, Guggino G, Stapylyton M, Badri MH, Solomon G, et al. IL-17 Inhibition in spondyloarthritis associates with subclinical gut microbiome perturbations and a distinctive IL-25-driven intestinal inflammation. *Arthritis Rheumatol* 2019; 72: 645–657.
 33. Smith G, Blackwell C, Nuki G. Faecal flora in spondyloarthropathy. *Br J Rheumatol* 1997; 36: 850–854.
 34. Navarro-Lopez V, Martinez-Andres A, Ramirez-Bosca A, Ruzafa-Costas B, Nunez-Delegido E, Carrion-Gutierrez MA, et al. Efficacy and safety of oral administration of a mixture of probiotic strains in patients with psoriasis: a randomized controlled clinical trial. *Acta Derm Venereol* 2019; 99: 1078–1084.
 35. Saunte DM, Mrowietz U, Puig L, Zachariae C. Candida infections in patients with psoriasis and psoriatic arthritis treated with interleukin-17 inhibitors and their practical management. *Br J Dermatol* 2017; 177: 47–62.
 36. Duarte PM, Nogueira CFP, Silva SM, Pannuti CM, Schey KC, Miranda TS. Impact of Smoking Cessation on Periodontal Tissues. *Int Dental J* 2021 Feb 27 [Online ahead of print].
 37. Gomes-Filho IS, Santos PNP, Cruz SS, Figueiredo ACMG, Trindade SC, Ladeia AM, et al. Periodontitis and its higher levels of severity are associated with the triglyceride/high density lipoprotein cholesterol (TG/HDL-C) ratio. *J Periodontol* 2021 Mar 10 [Online ahead of print].
 38. Vieira Colombo AP, Magalhães CB, Hartenbach FARR, Martins do Souto R, Maciel da Silva-Boghossian C. Periodontal-disease-associated biofilm: a reservoir for pathogens of medical importance. *Microb Pathog* 2016; 94: 27–34.
 39. Van Ende M, Wijnants S, Van Dijck P. Sugar sensing and signaling in *Candida albicans* and *Candida glabrata*. *Front Microbiol* 2019; 10: 99.
 40. Alam MT, Amos GCA, Murphy ARJ, Murch S, Wellington EMH, Arasaradnam RP. Microbial imbalance in inflammatory bowel disease patients at different taxonomic levels. *Gut Pathog* 2020; 12: 1.
 41. Matsuoka K, Kanai T. The gut microbiota and inflammatory bowel disease. *Semin Immunopathol* 2015; 37: 47–55.
 42. Koren O, Knights D, Gonzalez A, Waldron L, Segata N, Knight R, et al. A guide to enterotypes across the human body: meta-analysis of microbial community structures in human microbiome datasets. *PLoS Comput Biol* 2013; 9: e1002863.
 43. Turnbaugh PJ, Hamady M, Yatsunencko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. *Nature* 2009; 457: 480–484.
 44. Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. *Nature Rev Microbiol* 2020; 19: 55–71.
 45. Jensen P, Zachariae C, Christensen R, Geiker NRW, Schaadt BK, Stender S, et al. Effect of weight loss on the severity of psoriasis: a randomized clinical study. *JAMA Dermatol* 2013; 149: 795–801.
 46. Seregin SS, Golovchenko N, Schaf B, Chen J, Pudlo NA, Mitchell J, et al. NLRP6 protects *Il10*^{-/-} mice from colitis by limiting colonization of *Akkermansia muciniphila*. *Cell Rep* 2017; 19: 733–745.
 47. Ganesh BP, Klopffleisch R, Loh G, Blaut M. Commensal *Akkermansia muciniphila* exacerbates gut inflammation in *Salmonella typhimurium*-infected gnotobiotic mice. *PLoS One* 2013; 8: 74963.
 48. Ramirez-Boscá A, Navarro-López V, Martínez-Andrés A, Such J, Francés R, De La Parte J, et al. Identification of bacterial DNA in the peripheral blood of patients with active psoriasis. *JAMA Dermatol* 2015; 151: 670–671.
 49. Munch Roager H, Vogt JK, Kristensen M, Hansen LBS, Ibrügger S, Maerkedahl RB, et al. Whole grain-rich diet reduces body weight and systemic low-grade inflammation without inducing major changes of the gut microbiome: a randomised cross-over trial. *Gut* 2019; 68: 83–93.
 50. Trastoy B, Naegeli A, Anso I, Sjögren J, Guerin ME. Structural basis of mammalian mucin processing by the human gut O-glycopeptidase OgpA from *Akkermansia muciniphila*. *Nat Commun* 2020; 11: 1–14.
 51. Dao MC, Everard A, Aron-Wisnewsky J, Sokolovska N, Prifti E, Verger EO, et al. *Akkermansia muciniphila* and improved metabolic health during a dietary intervention in obesity: relationship with gut microbiome richness and ecology. *Gut* 2016; 65: 426–436.
 52. Duscha A, Gisevius B, Hirschberg S, Yissachar N, Stangl GI, Eilers E, et al. Propionic acid shapes the multiple sclerosis disease course by an immunomodulatory mechanism. *Cell* 2020; 180: 1067–1080.
 53. Venegas DP, De La Fuente MK, Landskron G, González MJ, Quera R, Dijkstra G, et al. Short chain fatty acids (SCFAs) mediated gut epithelial and immune regulation and its relevance for inflammatory bowel diseases. *Front Immunol* 2019; 10: 277.
 54. Schwarz A, Philippssen R, Schwarz T. Induction of regulatory t cells and correction of cytokine imbalance by short-chain fatty acids: implications for psoriasis therapy. *J Invest Dermatol* 2020; 141: 95–104.
 55. David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014; 505: 559–563.
 56. Sender R, Fuchs S, Milo R. Are we really vastly outnumbered? Revisiting the ratio of bacterial to host cells in humans. *Cell* 2016; 164: 337–340.
 57. Kristensen NB, Bryrup T, Allin KH, Nielsen T, Hansen TH, Pedersen O. Alterations in fecal microbiota composition by probiotic supplementation in healthy adults: a systematic review of randomized controlled trials. *Genome Med* 2016; 8: 52.
 58. Laursen MF, Laursen RP, Larnkjær A, Michaelsen KF, Bahl MI, Licht TR. Administration of two probiotic strains during early childhood does not affect the endogenous gut microbiota composition despite probiotic proliferation. *BMC Microbiol* 2017; 17: 175.
 59. Knight R, Vrbanac A, Taylor BC, Aksenov A, Callewaert C, Debelius J, et al. Best practices for analysing microbiomes. *Nature Rev Microbiol* 2018; 16: 410–422.
 60. Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, et al. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature* 2015; 528: 262–266.
 61. Imhann F, Bonder MJ, Vila AV, Fu J, Mujagic Z, Vork L, et al. Proton pump inhibitors affect the gut microbiome. *Gut* 2016; 65: 740–748.
 62. Donaldson GP, Lee SM, Mazmanian SK. Gut biogeography of the bacterial microbiota. *Nature Rev Microbiol* 2015; 14: 20–32.
 63. Graspentner S, Loeper N, Künzel S, Baines JF, Rupp J. Selection of validated hypervariable regions is crucial in 16S-based microbiota studies of the female genital tract. *Sci Rep* 2018; 8: 9678.
 64. Fouhy F, Clooney AG, Stanton C, Claesson MJ, Cotter PD. 16S rRNA gene sequencing of mock microbial populations-impact of DNA extraction method, primer choice and sequencing platform. *BMC Microbiol* 2016; 16: 123.
 65. Teng F, Darveekaran Nair SS, Zhu P, Li S, Huang S, Li X, et al. Impact of DNA extraction method and targeted 16S-rRNA hypervariable region on oral microbiota profiling. *Sci Rep* 2018; 8: 16321.
 66. Van Rossum T, Ferretti P, Maistrenko OM, Bork P. Diversity within species: interpreting strains in microbiomes. *Nature Rev Microbiol* 2020; 18: 491–506.
 67. Mills S, Shanahan F, Stanton C, Hill C, Coffey A, Ross RP. Movers and shakers. *Gut Microbes* 2013; 4: 4–16.