

Investigating the Role of I Kappa B Kinase ϵ in the Pathogenesis of Psoriasis

Isabella WEIMAR, Pernille OMMEN, Lars IVERSEN and Claus JOHANSEN*

Department of Dermatology, Aarhus University Hospital, Palle Juul-Jensens Boulevard 99, Aarhus N DK-8200, Denmark. *E-mail: Claus.Johansen@clin.au.dk

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Psoriasis is a common chronic inflammatory skin disease (1). Interleukin (IL)-17A has been identified as a key cytokine mediating the immunopathogenesis of psoriasis (2–4). However, the underlying molecular mechanism by which IL-17A mediates its psoriatic effects is unknown. A distinctive characteristic of psoriasis is the presence of neutrophils in the epidermis. Neutrophils are recruited to the epidermis by neutrophil-recruiting chemokines, such as C-X-C motif chemokine ligand (CXCL)1, CXCL5 and IL-8 produced by activated keratinocytes (5, 6). I kappa B kinase (IKK) ϵ is a protein encoded by the *IKBKE* gene. IKK ϵ has been demonstrated to play an important role in the recruitment of neutrophils in IL-17A-induced inflammation (7). By the use of IKK ϵ -deficient mice, it has been demonstrated that IKK ϵ is required for IL-17A-induced lung inflammation and, interestingly, IKK ϵ was found to play an important role in the recruitment of neutrophils (7). To date, the role of IKK ϵ in psoriasis remains unknown; however, the above-mentioned data suggest that IKK ϵ may play a role in the pathogenesis of psoriasis. The aim of the current study was to investigate the role of IKK ϵ in psoriasis.

MATERIALS AND METHODS (see Appendix S1¹)

RESULTS

In order to examine *IKBKE* mRNA expression, RNA was isolated from punch biopsies obtained from both lesional and non-lesional skin from 15 psoriatic patients and 6 patients with atopic dermatitis. Furthermore, RNA was isolated from 6 healthy controls. The mRNA expression of *IKBKE* was significantly increased in lesional psoriatic skin compared with non-lesional psoriatic skin from the same patient; an approximately 2-fold increase was observed. The expression of *IKBKE* in lesional psoriatic skin was also significantly higher compared with healthy controls (Fig. 1A). To determine whether the observed increase in *IKBKE* expression was specific to psoriasis or simply due to increased inflammation in the skin we also investigated *IKBKE* expression in atopic dermatitis. No changes were found in *IKBKE* expression between lesional and non-lesional atopic dermatitis skin (Fig. 1A). It was also found that the IKK ϵ protein level was increased in lesional psoriatic skin compared with non-lesional psoriatic skin from the same patient (Fig. 1B), supporting a potential role of IKK ϵ in the pathogenesis of psoriasis.

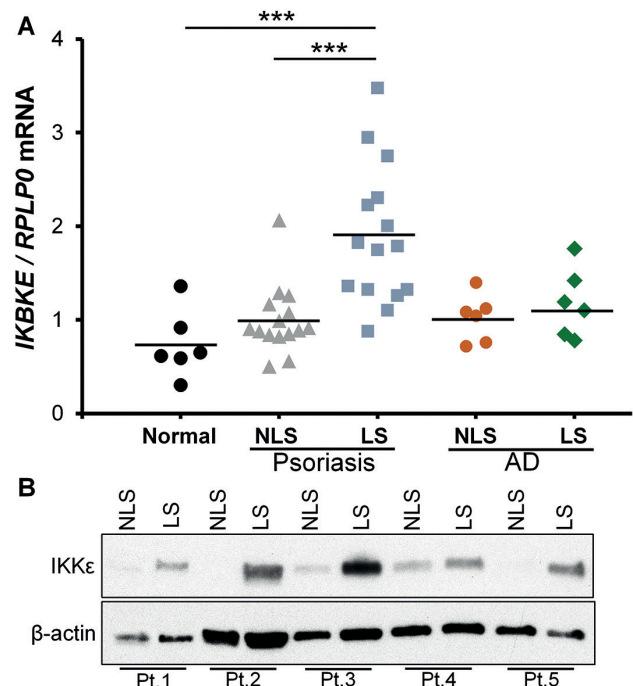


Fig. 1. Increased levels of I kappa B kinase ϵ (IKK ϵ) in lesional psoriatic skin. (A) RNA was extracted from lesional (LS) and non-lesional (NLS) skin biopsies from 15 patients with psoriasis, 6 patients with atopic dermatitis (AD) and from 6 healthy volunteers (Normal). *IKBKE* mRNA expression was analysed by quantitative PCR (qPCR). *RPLP0* mRNA was used for normalization. The relative fold induction is presented as scatter plots. Data were analysed with 1-way analysis of variance (ANOVA). *** $p < 0.001$. (B) IKK ϵ protein expression was examined in paired lesional (LS) and non-lesional (NLS) biopsies from 5 psoriatic patients (Pt.1–5) by Western blotting.

To characterize the role of IKK ϵ in the regulation of specific psoriasis-associated genes, siRNA was used to knockdown IKK ϵ . Transfection of human keratinocytes with IKK ϵ siRNA significantly reduced the mRNA and protein expression of *IKBKE*/IKK ϵ in tumour necrosis factor alpha (TNF α)- and/or IL-17A-stimulated cells compared with control siRNA transfected cells (Fig. S1¹). Notably, siRNA-mediated knockdown of IKK ϵ , led to a minor, but significant, reduction in *CCL20* mRNA expression after stimulation with IL-17A alone and with IL-17A in combination with TNF α , compared with control siRNA transfected cells (Fig. S2¹). Likewise, mRNA expression of *DEFB4* and *CXCL1* was significantly decreased in cells transfected with *IKBKE* siRNA, after combined stimulation with IL-17A and TNF α , but not when single stimulations were used (Fig. S2¹).

To further characterize the potential role of IKK ϵ in the pathogenesis of psoriasis the imiquimod-induced psori-

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asis-like skin inflammation model was used. First, *Ikkbe* mRNA expression in mice ears treated with imiquimod for 1, 3 and 5 days was investigated. Expression of *Ikkbe* mRNA was increased significantly at both days 3 and 5 in imiquimod-treated mice compared with vehicle-treated mice (Fig. S3¹). To assess the functional role of IKK ϵ in the imiquimod model, the model was applied on IKK ϵ -deficient mice. IKK ϵ deficiency in the mice was confirmed both by quantitative PCR (qPCR) and Western blotting (Fig. S4¹). Clinically, imiquimod-treated mice developed erythema and scaling of the skin. However, no differences were observed in the degree of erythema and scaling between IKK ϵ -deficient and wild-type mice treated with imiquimod (Fig. S5¹). Imiquimod treatment caused weight loss of approximately 15% and increased the ear thickness by approximately 130%; however, no differences in ear thickness were observed between imiquimod-treated wild-type and IKK ϵ -deficient mice (Fig. S6A, B¹). Haematoxylin and eosin (H&E) staining of skin biopsies revealed no histopathological differences between the 2 mice strains after treatment (Fig. S6C¹). Immunofluorescence staining revealed presence of neutrophils in skin sections after imiquimod treatment, however, no differences in the number of neutrophils between wild-type and IKK ϵ -deficient mice were observed (Fig. S6D¹). Finally, the expression of psoriasis-associated genes was examined in the ear biopsies. Although imiquimod treatment increased *Nfkbiz*, *Ccl20* and *Cxcl1* mRNA expression compared with vehicle-treated mice, no alterations in *Ccl20* and *Cxcl1* mRNA expression were observed between imiquimod-treated wild-type and IKK ϵ -deficient mice (Fig. S6E¹).

DISCUSSION

This study demonstrated an increased mRNA and protein expression of IKK ϵ in lesional psoriatic skin. The increase in IKK ϵ seemed to be specific for psoriasis and not just due to increased inflammation in the skin in general, because IKK ϵ expression was unaltered between lesional and non-lesional skin from patients with atopic dermatitis. Previous data have shown that the function of IKK ϵ in IL-17A-mediated signalling is linked with neutrophilia (7). IKK ϵ was demonstrated to play a role in IL-17A-induced airway inflammation by regulating the expression of neutrophil-mobilizing cytokines and chemokines, including CXCL1 and CXCL2 (7). Here, we also found IKK ϵ to be involved in the expression of CXCL1. However, only an involvement of IKK ϵ on the CXCL1 expression was observed when TNF α and IL-17A were given in combination, whereas no involvement was found by stimulation with IL-17A alone. The discrepancy between our data and that of Bulek et al. (7) could be because the role of IKK ϵ in IL-17A stimulation is cell-type specific.

To analyse how IKK ϵ was regulated *in vivo* during psoriasis development, we used the imiquimod-induced

psoriasis-like skin inflammation model, a well-described psoriasis mouse model (8). By comparing vehicle- and imiquimod-treated wild-type mice, we observed no increase in *Ikkbe* mRNA over time during imiquimod treatment. Likewise, we demonstrated only a minor, although significant, induction of *Ikkbe* mRNA upon imiquimod treatment at days 3 and 5. This is in contrast to what we observed in biopsies from psoriatic patients, where we demonstrated an almost 2-fold induction of *IKBKE* mRNA between lesional and non-lesional psoriatic skin. This could suggest that, in this specific psoriasis mouse model, or perhaps in mice in general, IKK ϵ does not play a major role in the induction of psoriasis-like skin inflammation. Although the imiquimod-induced psoriasis mouse model is dependent on IL-17A signalling (8, 9), it is possible that other cytokines might play a more prominent role in mice, and therefore mice might not represent the best model to study IL-17A downstream effects. This theory was also supported by the fact that we observed no difference between *Ikkbe* knockout mice and wild-type mice in their clinical appearance, bodyweight, ear thickness and expression of the inflammatory genes *Ccl20* and *Cxcl1* after imiquimod treatment. Surprisingly, IKK ϵ deficiency significantly increased the imiquimod-induced *Nfkbiz* gene expression, suggesting that IKK ϵ might have a protective role in this model.

Although we demonstrated increased expression of *Ikkbe* mRNA in lesional psoriatic skin and that IKK ϵ knockdown *in vitro* decreased IL-17A/TNF α -induced proinflammatory gene expression, this proinflammatory role of IKK ϵ in psoriasis was not supported by results from the imiquimod-induced psoriasis mouse model. Thus, further studies are needed in order to fully elucidate the role of IKK ϵ in psoriasis.

The authors have no conflicts of interest to declare.

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