

Testicular Cancer in Monozygotic Twin Brothers with Urticaria Pigmentosa

Tatiana PÉČOVÁ¹, Karolína VORČÁKOVÁ¹, Markéta ŽALIOVÁ², Tatiana BURJANIVOVÁ^{3,4}, Bibiana MALICHEROVÁ^{3,4}, Lukáš PLANK⁵, Jan TRKA², Klaudia PÉČOVÁ¹, Katarína ADAMICOVÁ⁵, Martin PÉČ⁶ and Juraj PÉČ^{1*}

Departments of ¹Dermato-Venereology, ²Pathology and ⁶Medical Biology, Jessenius Faculty of Medicine in Martin, Comenius University in Bratislava, Kollarova 2, 036 01 Martin, Slovakia, ³Childhood Leukaemia Investigation Prague (CLIP), Department of Paediatric Haematology and Oncology, ^{2nd} Faculty of Medicine, Charles University and University Hospital Motol, Prague, Czech Republic, Comenius University in Bratislava, Jessenius Faculty of Medicine in Martin, Biomedical Center Martin (JFM CU), ³Department of Molecular Biology and ⁴Division of Oncology JFM CU, Martin Slovakia. E-mail: jpec@jfmmed.uniba.sk

Accepted Dec 6, 2017; Epub ahead of print Dec 12, 2017

Urticaria pigmentosa (UP) is the most common variant of cutaneous mastocytosis. The occurrence of UP in twins is rare. We have described a case of monozygotic twin brothers, with the first signs of UP at the age of 4 months (1) and now present the development of their disease until 43 years of age, with the additional appearance of testicular cancer at age 30.

CASE REPORT

The UP lesions in both twins – diffuse macules affecting the whole body (Fig. 1), except the face – remained clinically unchanged during the 43 years of life apart from the short period of chemotherapy in one of the twins. In 2003, left orchiectomy was performed on one twin due to palpable testicular tumour, and in 2004, right orchiectomy was performed on the other due to the same reason. In both brothers the histology confirmed mixed germ cell tumour with the dominant component of embryonal carcinoma (more than 90% of the tumour) with a minor level of a yolk sac component and choriocarcinoma. Both twins were treated with 4 cycles of chemotherapy in monthly intervals (bleomycin, etoposide, cisplatin), and so far no recurrence of the tumour has been observed. In one twin, the UP lesions temporarily disappeared after the first cycle of chemotherapy, and reappeared after the last session, whereas in the second twin the UP lesions remained stable during the chemotherapy.

In both brothers, all routine biochemical parameters were within normal levels. The serum tryptase levels, 13.1 and 16.8 ng/ml, were both within reference values (below 20 ng/ml for adults – ImmunoCAP Tryptase test). The abdominal ultrasonography and chest X-ray were without any pathology.

The skin biopsy from both brothers had identical histological features of UP with an intact epidermis and a pleomorphic, mainly granulated mast cells with positive chloroacetate esterase, and CD117 – proto-oncogene *c-kit* in the upper corium. The bone

marrow trepanobiopsy in both brothers showed proportional representation of precursors of all 3 lines of hematopoiesis, the presence of trilinear maturation without proliferation of blasts with the presence of rare, dispersedly situated granulated mast cells (CD117⁺, CD25⁻), without infiltration of mast cell populations, whereas the skeletal scintigraphy was negative (with Tc-MDP methylene diphosphonate as imaging agent).

Whole blood samples and saliva were collected from the twins. DNA was isolated by using DNeasy Blood and Tissue Kit (Qiagen). First we worked with DNA isolated from the whole blood, where Sanger sequencing of the entire coding region of the proto-oncogene *KIT* was performed (2). Since Sanger sequencing of *c-kit* gene did not reveal any pathogenic mutation, we decided to analyse the whole exome, where we found *D816V* of the *KIT* gene in both samples. However, the mutation rate was very low, which is why Sanger sequencing did not reveal any mutation. Assuming that the twins originate from a mosaic embryo, we isolated DNA from the saliva for further examination. Allele-specific PCR (AS-PCR) for the *KIT D816V* were performed in patients' saliva samples (3). We found *D816V* mutation in both twins (Fig. S1¹).

Sequencing libraries were prepared from DNA using Agilent SureSelectXT HumanAllExon V5 kit according to the manufacturer's instructions (Agilent, Technologies, USA). High throughput sequencing (2x75 cycles) was performed on NextSeq500 using High Output Kit (Illumina, USA). Read pairs were aligned to the human genome reference (hg19) using Burrows-Wheeler aligner and further processed by Picard tools (<http://broadinstitute.github.io/picard/>) (4). Variant calling was performed using VarScan (5). Regions of interest were visually inspected in Integrative Genomics Viewer (IGV) (6). Part of the exon 17 (Chr4: 55599271-55599370; hg19) of the *KIT* gene (NM_000222) surrounding codon D816 was amplified by single-round PCR using primer pairs listed in Table S1¹.

¹<https://doi.org/10.2340/00015555-2861>

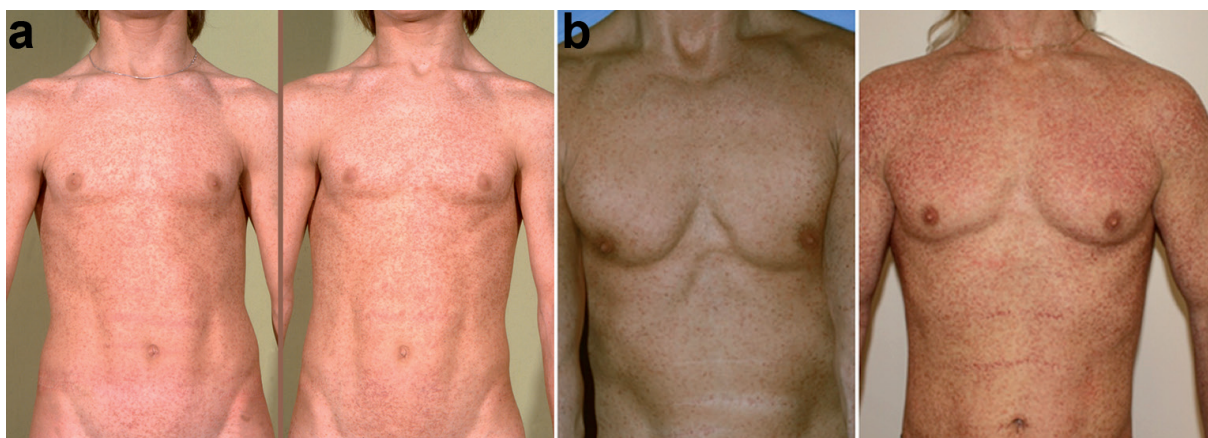


Fig. 1. The brothers' skin phenotypes at the age of 16 years (a) and 43 years (b), respectively.

The resulting indexed libraries were sequenced on Ion Torrent PGM using 400bp chemistry according to the manufacturer's instructions (Life Technologies, USA). Fastq files were processed from raw data and reads were mapped to hg19 using Torrent Suite software (Life Technologies). Variant calling was performed using Variant Caller plugin in Torrent Suite software (parameter settings: somatic variant frequency, low stringency). Mapped reads were visually inspected in IGV. Codon D816 was covered by >25,000 reads in both patient samples peripheral blood samples and by >10,000 reads in saliva samples. To distinguish mutations from the potential PCR and/or sequencing errors, error rate at the respective position was analyzed in two buffy coat samples from healthy individuals. The frequency of A>T error at chr4: 55599321 was 0.04% and 0.05%, respectively (position coverage >22,000 reads).

Using whole exome sequencing (WES), the KIT gene mutation D816V was identified at a relatively low frequency in peripheral blood of first twin (variant allele frequency 9%; 5 mutated out of total 56 reads). Although no KIT mutation was found in the second twin, a visual inspection of mapped reads in IGV revealed KIT gene mutation identical to that found in first twin in one out of total 47 reads. To validate these findings deep amplicon sequencing of the respective KIT gene region was performed using the same DNA samples. In agreement with WES results, D816V mutation was detected at low level in peripheral blood of both brothers with variant allele frequency 9% in first twin and 3% in second twin. Subsequent deep sequencing analysis revealed D816V mutation also in saliva samples: 5% and 2% in first and second twin, respectively.

DISCUSSION

Familial cutaneous mastocytosis – especially in monozygotic twins – occurs very rarely (7) and is considered as genetically determined, perhaps autosomal dominant disease (8). However, Sato-Matsumura et al. (9) did not find any gene mutations at exon 11 or 17 *c-kit* in monozygotic twins with UP. Mastocytosis in childhood is defined as disease starting before the age of 15 years (8), usually affecting only the skin, perhaps as a clonal proliferation of benign prognosis (10), in contrast to the beginning of the disease after age 15 with chronic progression, development of haematological malignancies (11), as well as induction of potentially oncogenic somatic *c-KIT* mutations (12, 13). The serum tryptase levels generally correlate with the proliferation and bone marrow infiltration of mast cells (10, 11). Elevation in serum tryptase above 20 ng/dl is an indication for bone marrow trepanobiopsy.

In the described monozygotic twins, the skin lesions were *KITD816V* negative, there were no mast cell infiltration of the bone marrow, and serum tryptase was below the reference level. However, the *D816V* mutation was detected in low levels in peripheral blood as well as saliva samples. The *KIT D816V* mutation does not necessarily correlate with clinical manifestations, as investigated in indolent systemic mastocytosis (14). However, its detection may prove very useful in patients with systemic mastocytosis but very low mast cell burden, and who do not meet WHO 2008 histological criteria for bone marrow involvement (15). Serum tryptase levels, on the other hand, correlate with the burden of *D816V* mutation, making it a disease marker in systemic mastocytosis (16).

The twins we describe presumably originated from a mosaic embryo and as such presented no systemic signs of mastocytosis. The clinical picture correlates to low level *KIT D816V* mutation in peripheral blood and saliva. This probably represent a mild systemic form of mastocytosis with some predisposition to future systemic complications. Whether or not testicular cancer in the twins is part of this predisposition or represents an unrelated genetic trait remains to be elucidated.

REFERENCES

1. Pec J, Palencarova E, Malisova S, Dobrota D, Hajtman A, Pec M, Lepej J. Urticaria pigmentosa in identical twins. *Acta Derm Venereol* 1995; 75: 244–252.
2. De Melo Campos P, Machado-Neto JA, Scopim-Ribeiro R, Visconte V, Tabarroki A, Duarte AS, et al. Familial systemic mastocytosis with germline KIT K509I mutation is sensitive to treatment with imatinib, dasatinib and PKC412. *Leuk Res* 2014; 38: 1245–1251.
3. Corless CL, Harrell P, Lacouture M, Bainbridge T, Le C, Gatter K, et al. Allele-specific polymerase chain reaction for the imatinib-resistant KIT D816V and D816F mutations in mastocytosis and acute myelogenous leukemia. *J Mol Diagn* 2006; 8: 604–612.
4. Li H, Durbin R. Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics* 2010; 26: 589–595.
5. Koboldt DC, Zhang Q, Larson DE, Shen D, McLellan MD, Lin L, et al. VarScan 2: somatic mutation and copy number alteration discovery in cancer by exome sequencing. *Genome Res* 2012; 22: 568–576.
6. Robinson JT, Thorvaldsdóttir H, Winckler W, Guttman M, Lander ES, Getz G, Mesirov JP. Integrative Genomics Viewer. *Nature Biotechnology* 2011; 29: 24–26.
7. De La Sotta P, Romero WA, Kramer D, Cárdenas C, Gonzáles S. Cutaneous mastocytosis in twins: Multiple mastocytomas and urticaria pigmentosa in two pairs of monozygotic twins. *Pediatr Dermatol* 2011; 28: 585–587.
8. Heide R, Tank B, Oranje AP. Mastocytosis in childhood. *Pediatr Dermatol* 2002; 19: 375–381.
9. Sato-Matsumura KC, Matsumura T, Koizumi H, Sato H, Nagashima K, Ohkawara A. Analysis of c-kit exon 11 and exon 17 of urticaria pigmentosa occurred in monozygotic twin sisters. *Br J Dermatol* 1999; 140: 1130–1132.
10. Kiszewski AE, Durán-Mckinster C, Orozco-Covarrubias L, Gutiérrez-Castrellón P, Ruiz-Maldonado R. Cutaneous mastocytosis in children: a clinical analysis of 71 cases. *J Eur Acad Dermatol Venereol* 2004; 18: 285 D2290.
11. Shaffer HC, Parsons DJ, Peden DB, Morrell D. Recurrent syncope and anaphylaxis as presentation of systemic mastocytosis in a pediatric patient. Case report and literature review. *J Am Acad Dermatol* 2006; 54: 210–213.
12. Zanotti R, Simioni L, Garcia-Montero AC, Perbellini O, Bonadonna P, Caruso B, et al. Somatic D816V KIT mutation in a case of adult-onset familial mastocytosis. *J Allergy Clin Immunol* 2013; 131: 605–607.
13. Broesby-Olsen S, Kristensen TK, Møller MB, Bindslev-Jensen C, Vestergaard H. Adult-onset systemic mastocytosis in monozygotic twins with KIT D816V and JAK2 V617F mutations. *J Allergy Clin Immunol* 2012; 130: 806–808.
14. Broesby-Olsen S, Kristensen T, Vestergaard H, Brixen K, Møller MB, Bindslev-Jensen C. KIT D816V mutation burden does not correlate to clinical manifestations of indolent systemic mastocytosis. *J Allergy Clin Immunol* 2013; 132: 723–728.
15. De Matteis G, Zanotti R, Colarossi S, De Benedittis C, Garcia-Montero A, Bonifacio M, et al. The impact of sensitive KIT D816V detection on recognition of indolent Systemic Mastocytosis. *Leuk Res* 2015; 39: 273–278.
16. Kristensen T, Broesby-Olsen S, Vestergaard H, Bindslev-Jensen C, Møller MB. Serum tryptase correlates with the KIT D816V mutation burden in adults with indolent systemic mastocytosis. *Eur J Haematol* 2013; 91: 106–111.