INVESTIGATIVE REPORT

Platelet-derived Growth Factor Receptor Alpha Gene Mutations in Vitiligo Vulgaris

Shengxin XU^{1,2}, Youwen ZHOU¹⁻³, Sen YANG^{1,2}, Yunqing REN^{1,2}, Chi ZHANG^{1,2}, Cheng QUAN^{1,2}, Min GAO^{1,2}, Caifeng HE^{1,2}, Hui CHEN^{1,2}, Jianyen HHAN^{1,2}, Jianjun CHEN^{1,2}, Yanhua LIANG^{1,2}, Jianqiang YANG^{1,2}, Liangdan SUN^{1,2}, Xianyong YIN^{1,2}, Jianjun LIU^{1,2} and Xuejun ZHANG^{1,2}

¹Institute of Dermatology & Department of Dermatology, The First Affiliated Hospital, Anhui Medical University, ²The Key Laboratory of Gene Resource Utilization for Severe Genetic Diseases, Ministry of Education and Anhui Province, Hefei, Anhui, China, and ³Chieng Genomics Center and Laboratory of Predictive Medicine and Therapeutics, Vancouver Coastal Health Research Institute, and Department of Dermatology and Skin Science, University of British Columbia, Vancouver, Canada

Vitiligo vulgaris is an acquired depigmenting disorder resulting from the loss of melanocytes in the skin. Though several putative susceptibility loci of vitiligo have been identified in different populations, the pathogenesis of the disease remains poorly understood. Through genetic linkage analysis of a large Chinese family cohort of vitiligo, we identified a vitiligo linkage locus AIS4 within chromosome 4q12-q21, a region containing several possible candidate genes, including the plateletderived growth factor receptor alpha (PDGFRA) gene. We postulated that PDGFR mutations may be linked with vitiligo. To test this hypothesis, we performed DNA sequencing on this gene in 143 multiplex families with familial vitiligo vulgaris, 480 patients with sporadic vitiligo vulgaris, and 480 healthy subjects. Mutations were found in 3.5% of familial vitiligo cases, which is significantly higher than for the general population (0.42%, p=0.008, Fisher's exact test), and possibly higher than in sporadic vitiligo patients (1.0%, p=0.053). To our knowledge, this is the first observation that PDGFRA mutations are linked with familial vitiligo vulgaris. Key words: PDGFRA; gene; mutation; vitiligo.

(Accepted December 7, 2009.)

Acta Derm Venereol 2010; 90: 131-135.

Xuejun Zhang, Institute of Dermatology, Anhui Medical University, 69 Meishan Road, Hefei, Anhui 230032, China. E-mail: ayzxj@vip.sina.com

Vitiligo vulgaris (MIM193200) is an acquired, noncontagious disorder in which progressive loss of viable melanocytes results in patchy pigmentation of the skin, hair and oral mucosa (1). It affects all populations worldwide, with diverse prevalence rates ranging from 0.1% to 2% among different geographical regions and ethnic groups (1). In some cases the disorder is associated with other syndromes, such as autoimmune diseases (2, 3). At present, it is generally accepted that the pathogenesis of the disease is multifactorial and genetically heterogeneous (4). Mutations or polymorphisms in several genes have been associated with development of vitiligo (5–8). However, together they account for only a small proportion of the genetic susceptibility for vitiligo. Recent gene mapping studies have identified several major susceptibility loci for vitiligo (9), including evidence for a major vitiligo susceptibility locus on chromosome 4q12-q21 (*AIS4*) in the Chinese population (10). However, the genes responsible have not been identified.

The platelet-derived growth factor receptor alpha (*PDGFRA*) gene is a proto-oncogene that maps to 4q12. It belongs to the human type III family of transmembrane receptors, with an intrinsic tyrosine kinase component. The PDGFRA protein has been shown to be important for several cellular and tissue processes, such as proliferation, apoptosis, chemotaxis, melanogenesis, hematopoiesis and gametogenesis (11). Several reports have documented PDGFRA in regulation of pigmentation. First, a deletion encompassing the PDGFRA gene, found in Patch (Ph) mutant mice, initially studied by Gruneberg & Truslove (12), is associated with defective melanocyte migration in heterozygotes leading to the development of white patches on the trunk. Secondly, several reports have documented that vitiligo is severely exacerbated by protein tyrosine kinase inhibitor therapy (13). Thirdly, platelet-derived growth factor (PDGF) has been shown to be important for the differentiation and survival of melanocytes during embryonic development (14). These reports lead us to speculate that the PDGFRA gene may be a candidate susceptibility gene of vitiligo mapped to the region of 4q12, and prompted us to sequence the coding region of this gene in a large number of familial and sporadic vitiligo cases using healthy individuals as the controls. Our results show that a small, but not insignificant, number of patients with familial vitiligo carry mutations in the PDGFRA gene, significantly more than the general population, supporting a role for this gene in the development of at least some cases of vitiligo.

MATERIALS AND METHODS

Subjects

The subjects studied were recruited from three different populations: the vitiligo families, sporadic cases, and controls. A total of 143 multiplex vitiligo Chinese Han families, including the 106 pedigrees used in our previous genome-wide linkage study (10), were recruited from the Dermatology Department of the Anhui Medical University and the Vitiligo Clinic of the Railway Hospital, Xiangfan, Hubei, China. Each family had at least two siblings with vitiligo. We first sequenced one case in each family (the family proband). We then determined the mutation status at the mutation site for the rest of the members (both affected and unaffected) of the probands carrying the mutations. Furthermore, the sites of the PDGFRA gene carrying the mutations were sequenced in 480 sporadic vitiligo cases and an equal number of controls, recruited from the same institutions.

The diagnosis of vitiligo was made on the basis of patient's history and typical clinical features consisting of discrete, wellcircumscribed and depigmented patches. Phenotypes were determined by history, lesion maps, physical examination and/or photographs. Any individual whose phenotype was questionable was excluded from this study, in particular atypical lesion distribution and congenital depigmentation. Only patients with clear signs of acquired patches on the extremities, trunk, genitalia, central face, or other areas were classified as affected. All the participating individuals provided informed written consent. The study was approved by the ethics committee of Anhui Medical University and was conducted according to Declaration of Helsinki principles.

Genetic analysis

Genomic DNA was extracted from peripheral blood leukocytes using a standard procedure (15). We designed primers flanking the entire coding sequence, exon-intron junctions, promoter regions and 5'-and 3'-untranslated regions of PDGFRA using the web-based version of the Primer 3.0 program (http://frodo. wi.mit.edu/primer3/). Primer sequences were available on request. A polymerase chain reaction (PCR) was performed in 10 µl reaction volume containing 20 ng of genomic DNA, 0.3 mM dNTPs, 0.3 µM of each primer, 3.0 mM MgCl2 and 0.1 units of Hotstar® Tag DNA polymerase (Qiagen, Hilden, Germany). The PCR conditions were: Hotstar[®] Tag activation at 95°C for 15 min, followed by 40 cycles, each having denaturation at 94°C for 40 s, annealing at 57°C for 60 s and extension at 72°C for 55 s, except that in the first 14 cycles the annealing temperature decreased

G

Т

c.3154A>T (p.T1052S)



Fig. 1. Representative sequencing read-outs showing the five PDGFRA mutations in familial vitiligo.

Table I. Five rare PDGFRA mutations in three different populations (the vitiligo families, sporadic cases, and controls)

PDGFRA	Protein		Frequency			
mutations	designations	Position	Families	Sporadic	Control	
c.367-3C>T	a	Intron 4	0/143	1/480	0/480	
c.418G>T	p.V140L	Exon 4	1/143	0/480	0/480	
c.2986G>A	p.E996K	Exon 22	1/143	0/480	0/480	
c.3098A>T	p.D1033V	Exon 22	2/143	4/480	2/480	
c.3154A>T	p.T1052S	Exon 23	1/143	0/480	0/480	
Total			5/143	5/480	2/480	
			(3.5%)	(1%)	(0.4%)	

^aThis mutation is located in the acceptor splice site and could prevent proper splicing of the transcript.

from 64°C to 57°C by 0.5°C per cycle, and the final extension was 72°C for 10 min. After the amplification, products were purified using a QIAquick PCR Purification Kit (Qiagen) and directly sequenced on ABI PRISM® 3730 automated sequencer (Applied Biosystems). Sequence comparisons and analysis were performed using Phred-Phrap-Consed Version 12.0 program. The A of the ATG of the initiator Met codon is denoted as nucleotide +1. PDGFRA GenBank sequences used: NM 006206 (mRNA); NP 006197 (protein).

RESULTS

We first screened for sequence variants of PDGFRA in genomic DNA samples from one affected individual (proband) from each of 143 families affected with vitiligo. Four non-synonymous mutations (c.418G>T, p.V140L; c.2986G>A, p.E996K; c.3098A>T, p.D1033V; and c.3154A>T. p.T1052S) were identified in the PDGFRA gene (Table I and Fig. 1). The four mutations were found in a total of five families. None of these sequence variants have been listed in the National Center for Biotechnology Information (NCBI) or Celera single nucleotide poly-

morphism (SNP) databases.

To investigate whether these mutations co-segregated with vitiligo, we analyzed all available DNA samples from all members of the above five families affected with vitiligo. The pedigrees of the five families showing the mutation status and phenotypic information for each member are shown in Fig. 2. The missense mutation c.3098A>T (p.D1033V) was observed in two different families, and the remaining mutations affected single families only. In every case, the mutations co-segregated with the vitiligo phenotypes within the affected families.

We further evaluated the association between these mutations and vitiligo by screening 480

С A Т TG A G С G G ТТ С



sporadic cases and 480 controls. It was found that 5 of 480 (1%) sporadic vitiligo vulgaris patients carry mutations in the *PDGFRA* gene, compared with 2 of 480 (0.42%) of control unaffected subjects (Table I). Of note, *PDGFR* mutation rate in familial vitiligo cases (3.5%) is significantly higher than in the control population (0.42%, p=0.008042, Fisher's exact test), probably higher than in sporadic vitiligo patients (1.0%, p=0.053306), although the mutation rates between sporadic vitiligo and control populations were not significantly different (p=0.225704). More clinical details of the patients are presented in Table II.

DISCUSSION

In the present study, we have identified five novel mutations in the *PDGFRA* gene associated with vitiligo. To our knowledge, this is the first time that *PDGFRA*

Fig. 2. Pedigrees representing the five generalized vitiligo families with *PDGFRA* mutations identified in this study. The probands are indicated by arrows. Black symbols denote affected individuals and unfilled symbols denote the unaffected individuals. Under each symbol, the first line corresponds to the current age of subjects; the second line shows the age at diagnosis of overt generalized vitiligo (when known). The third line shows the genotype at *PDGFRA* gene: N: normal allele; M: mutant allele.

has been implicated as a predisposing gene for development of vitiligo.

PDGFRA is a gene that spans 23 exons. It encodes a transmembrane protein composed of five immunoglobulin-like domains in the extracellular region, a transmembrane domain, an adenosine-triphosphate binding site, and a hydrophilic kinase insert domain in the intracellular portion (16). Upon binding to the extracellular immunoglobulin-like domains with the ligand (PDGF), PDGFR proteins form dimers, leading to the activation of the intrinsic tyrosine kinase activity. Four of five PDG-FRA missense mutations described here, the p.V140L, p.E996K, p.D1033V, and p.T1052S, are linked to an autosomal-dominant form of vitiligo (Fig. 3). The p.V140L point mutation is localized to exon 4 of PDGFRA, which encodes parts of the extracellular domain of PDGFRA, which may modulate ligand-binding and dimerization, thereby indirectly influence tyrosine kinase activity.

Table II. Clinical features of patients with platelet-derived growth factor receptor alpha mutations

Patient ID		Sex/age (years)	Onset (years)	Clinical type	Course	Severity	Other autoimmune disease	Mutation
Family F-039	II1 II2	F/31 M/40	19 33	Universal Universal	Progressive Progressive	Moderate Moderate	Not observed Not observed	c.3098A>T
Family F-060	II1 II2	F/38 F/34	19 16	Localized Localized	Stable Stable	Mild Mild	Chronic urticaria Not observed	c.3098A>T
Family F-149	II1 II2 II3	F/19 F/17 F/16	10 11 7	Localized Generalized Generalized	Decubation Decubation Decubation	Mild Mild Mild	Not observed Not observed Not observed	c.3154A>T
Family F-095	II1 II2	M/31 M/34	14 16	Localized Acrofacial	Decubation Stable	Mild Mild	Not observed Not observed	c. 418G>T
Family F-049	II1 II4 III2	M/83 M/86 F/52	3 20 49	Generalized Generalized Localized	Stable Stable Progressive	Moderate Moderate Mild	Not observed Not observed Diabetes mellitus	c.2986G>A
Sporadic Sporadic	case307 case156	F/20 M/11	20 8	Localized Localized	Stable Progressive	Mild Mild	Not observed Not observed	c.367-3C>T c.3098A>T
Sporadic Sporadic Sporadic	case191 case197 case389	M/24 M/15 M/11	20 13 9	Localized Localized Generalized	Stable Stable Progressive	Mild Mild Mild	Not observed Not observed Not observed	c.3098A>T c.3098A>T c.3098A>T



Fig. 3. Location of the mutations in the PDGFRA protein. The locations of the mutations identified in this study are shown on the left. The results from protein alignment of multiple genomes are shown on the right. Ig: immunoglobulinlike domains; TM: transmembrane domain; JM: juxtamembrane domain; KI: kinase insert; K1: kinase domain I; K2: kinase domain II; CT: C-terminal tail; Ma: Macaca fascicularis (crab-eating macaque); m: Mus musculus (mouse); r: Rattus norvegicus (Norway rat); d: Canis familiaris (dog); z: Danio rerio (zebrafish).

The other three missense mutations (p.E996K, p.D1033V, and p.T1052S) are localized to the Cterminal tail of PDGFRA. The exact function of these missense mutations found in this study is unknown. It is known that the C-terminal region from residue 977 to 1024 of PDGFRA is required for ligand-dependent focus formation, and that the tyrosine residues 988 and 1018 located within this domain constitute the major binding site for phospholipase C- γ (PLC- γ) (17). Therefore, the three mutations identified in this study may interfere with the catalytic actions of the PDGFRA, although more studies are needed to verify this possibility. Finally, the c.367-3 C>T mutation was identified in one sporadic patient. This mutation is located in the splice acceptor site and may result in mis-splicing of the PDGFRA transcript.

PDGFR is a receptor tyrosine kinase that is known to be required for melanocyte development. Several lines of clinical observation suggest that proper function of receptor tyrosine kinase receptor signaling pathways are required for melanocyte viability in mature adult human skin: First, Legros et al. (13) have observed a marked progression of a vitiligo after treatment with tyrosine kinase inhibitors in a patient with stable vitiligo

Acta Derm Venereol 90

for many years. Secondly, several cases of vitiliginous depigmentation occurring after treatment with new tyrosine kinase inhibitors (STI-571 and SU11428) have been reported (18–21). Imatinib mesilate is a selective inhibitor of several tyrosine kinases, such as PDGFRA and KIT, another melanocyte receptor shown to be required for melanocyte survival. Previously, most of the explanations for the observed effect of tyrosine kinase inhibition on melanocyte survival focused on the role of KIT receptor. However, our results suggest that PDGFR, which also is inhibited by the same inhibitors, could also be responsible, or contribute to the death of melanocytes in patients undergoing tyrosinase kinase inhibitor therapies.

It should be pointed out that the number of pedigrees containing *PDGFRA* mutations is low overall, and thus cannot explain the major linkage signal observed in the initial genome-wide linkage studies (10). Therefore, additional candidate genes in the 4q12 region, such as *KIT* gene, may still exist in addition to the *PDGFRA* gene. Detailed investigation of *KIT* gene in the familial vitiligo cases is in progress, and will be reported elsewhere.

Vitiligo is a complex disease that is strongly influenced by both genetic and environmental factors. While the pathogenesis is unknown, multiple hypotheses have been proposed to explain the clinical observations. The most commonly referred pathogenesis mechanisms include: genetic hypothesis (9) or intrinsic hypothesis (22, 23), autoimmune hypothesis (24), autocytotoxic hypothesis (25) and neural hypothesis (26). Our results are consistent with the genetic/intrinsic hypothesis. Two general models of complex diseases have been proposed. One is the common disease-common variant model, which holds that genetic susceptibility to common diseases is conferred primarily by alleles that are common in the population and have modest phenotypic effects (27, 28). This model is supported by a number of well-validated examples (29-32). The alternative model is that susceptibility to common diseases is the result of multiple rare alleles with large phenotypic effects (common disease-rare variant model). Although individually rare, these alleles may be collectively common in the population (33–35).

In summary, we have discovered that mutations of *PDGFRA* gene are associated with vitiligo development in a small number of cases of familial and sporadic vitiligo in China. This observation lends support to the genetic/intrinsic hypothesis of vitiligo pathogenesis. Specifically abnormalities in the genes involved in melanocyte survival/viability may lead to melanocyte death, causing vitiligo.

ACKNOWLEDGEMENTS

We thank all the participants of the study. This study was supported in part by the Key Project of National Natural Science Foundation (No. 30530670 to XJZ), the Cooperation Project of Chinese Key National Natural Science Foundation for Overseas Youth (No. 30628021 to YZ and XJZ), Anhui Provincial Special Scientific Program (2007-7, to XJZ), Constructive Foundation from the Key Lab of Gene Resources Utilization for Severe Genetic Disease, Anhui Province and Ministry of Education, China (to XJZ). We thank Dr Sancy Leachman for her kind assistance in preparation of this manuscript.

REFERENCES

- Hann SK, Nordlund J. Vitiligo: a comprehensive monograph on basic and clinical science. New York: Blackwell Science, 2000.
- Hofmann UB, Brocker EB, Hamm H. Simultaneous onset of segmental vitiligo and a halo surrounding a congenital melanocytic naevus. Acta Derm Venereol 2009; 89: 402–406.
- 3. Yalcin B, Tamer E, Gur G, Oztas P, Polat MU, Alli N. Neurofibromatosis 1/Noonan syndrome associated with Hashimoto's thyroiditis and vitiligo. Acta Derm Venereol 2006; 86: 80–81.
- Hafez M, Sharaf L, Abd el-Nabi SM. The genetics of vitiligo. Acta Derm Venereol 1983; 63: 249–251.
- Pehlivan S, Ozkinay F, Alper S, Onay H, Yuksel E, Pehlivan M, et al. Association between IL4 (-590), ACE (I)/(D), CCR5 (Delta32), CTLA4 (+49) and IL1-RN (VNTR in intron 2) gene polymorphisms and vitiligo. Eur J Dermatol 2009; 19: 126–128.
- Li M, Gao Y, Li C, Liu L, Li K, Gao L, et al. Association of COX2 functional polymorphisms and the risk of vitiligo in Chinese populations. J Dermatol Sci 2009; 53: 176–181.
- Birlea SA, Laberge GS, Procopciuc LM, Fain PR, Spritz RA. CTLA4 and generalized vitiligo: two genetic association studies and a meta-analysis of published data. Pigment Cell Melanoma Res 2009; 22: 230–234.
- Ren Y, Yang S, Xu S, Gao M, Huang W, Gao T, et al. Genetic variation of promoter sequence modulates XBP1 expression and genetic risk for vitiligo. PLoS Genet 2009; 5: e1000523.
- 9. Zhang XJ, Chen JJ, Liu JB. The genetic concept of vitiligo. J Dermatol Sci 2005; 39: 137–146.
- Chen JJ, Huang W, Gui JP, Yang S, Zhou FS, Xiong QG, et al. A novel linkage to generalized vitiligo on 4q13-q21 identified in a genomewide linkage analysis of Chinese families. Am J Hum Genet 2005; 76: 1057–1065.
- Mol CD, Lim KB, Sridhar V, Zou H, Chien EY, Sang BC, et al. Structure of a c-kit product complex reveals the basis for kinase transactivation. J Biol Chem 2003; 278: 31461–31464.
- 12. Gruneberg H, Truslove GM. Two closely linked genes in the mouse. Genet Res 1960; 1: 69–90.
- Legros L, Cassuto JP, Ortonne JP. Imatinib mesilate (Glivec): a systemic depigmenting agent for extensive vitiligo? Br J Dermatol 2005; 153: 691–692.
- Adameyko I, Lallemend F, Aquino JB, Pereira JA, Topilko P, Muller T, et al. Schwann cell precursors from nerve innervation are a cellular origin of melanocytes in skin. Cell 2009; 139: 366–379.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res 1988; 16: 1215.
- Kawagishi J, Kumabe T, Yoshimoto T, Yamamoto T. Structure, organization, and transcription units of the human alpha-platelet-derived growth factor receptor gene, PDG-FRA. Genomics 1995; 30: 224–232.
- 17. Yu JC, Li W, Wang LM, Uren A, Pierce JH, Heidaran MA. Differential requirement of a motif within the carboxyl-terminal domain of alpha-platelet-derived growth

factor (alpha PDGF) receptor for PDGF focus forming activity chemotaxis, or growth. J Biol Chem 1995; 270: 7033–7036.

- Raanani P, Goldman JM, Ben-Bassat I. Challenges in oncology. Case 3. Depigmentation in a chronic myeloid leukemia patient treated with STI-571. J Clin Oncol 2002; 20: 869–870.
- Hasan S, Dinh K, Lombardo F, Dawkins F, Kark J. Hypopigmentation in an African patient treated with imatinib mesylate: a case report. J Natl Med Assoc 2003; 95: 722–724.
- Tsao AS, Kantarjian H, Cortes J, O'Brien S, Talpaz M. Imatinib mesylate causes hypopigmentation in the skin. Cancer 2003; 98: 2483–2487.
- Brazzelli V, Roveda E, Prestinari F, Barbagallo T, Bellani E, Trevisan V, et al. Vitiligo-like lesions and diffuse lightening of the skin in a pediatric patient treated with imatinib mesylate: a noninvasive colorimetric assessment. Pediatr Dermatol 2006; 23: 175–178.
- 22. Boissy RE, Liu YY, Medrano EE, Nordlund JJ. Structural aberration of the rough endoplasmic reticulum and melanosome compartmentalization in long-term cultures of melanocytes from vitiligo patients. J Invest Dermatol 1991; 97: 395–404.
- Boissy RE, Beato KE, Nordlund JJ. Dilated rough endoplasmic reticulum and premature death in melanocytes cultured from the vitiligo mouse. Am J Pathol 1991; 138: 1511–1525.
- 24. Wankowicz-Kalinska A, Le Poole C, van den Wijngaard R, Storkus WJ, Das PK. Melanocyte-specific immune response in melanoma and vitiligo: two faces of the same coin? Pigment Cell Res 2003; 16: 254–260.
- Lerner AB, Nordlund JJ. Vitiligo: the loss of pigment in skin, hair and eyes. J Dermatol 1978; 5: 1–8.
- Koga M. Vitiligo: a new classification and therapy. Br J Dermatol 1977; 97: 255–261.
- Chakravarti A. Population genetics making sense out of sequence. Nat Genet 1999; 21: 56–60.
- Lander ES. The new genomics: global views of biology. Science 1996; 274: 536–539.
- 29. Zhang XJ, Huang W, Yang S, Sun LD, Zhang FY, Zhu QX, et al. Psoriasis genome-wide association study identifies susceptibility variants within LCE gene cluster at 1q21. Nat Genet 2009; 41: 205–210.
- Cargill M, Schrodi SJ, Chang M, Garcia VE, Brandon R, Callis KP, et al. A large-scale genetic association study confirms IL12B and leads to the identification of IL23R as psoriasis-risk genes. Am J Hum Genet 2007; 80: 273–290.
- 31. Musone SL, Taylor KE, Lu TT, Nititham J, Ferreira RC, Ortmann W, et al. Multiple polymorphisms in the TNFAIP3 region are independently associated with systemic lupus erythematosus. Nat Genet 2008; 40: 1062–1064.
- 32. Graham RR, Cotsapas C, Davies L, Hackett R, Lessard CJ, Leon JM, et al. Genetic variants near TNFAIP3 on 6q23 are associated with systemic lupus erythematosus. Nat Genet 2008; 40: 1059–1061.
- 33. Lee-Kirsch MA, Gong M, Chowdhury D, Senenko L, Engel K, Lee YA, et al. Mutations in the gene encoding the 3'-5' DNA exonuclease TREX1 are associated with systemic lupus erythematosus. Nat Genet 2007; 39: 1065–1067.
- Polychronakos C. Common and rare alleles as causes of complex phenotypes. Curr Atheroscler Reports 2008; 10: 194–200.
- Yang C-F, Hwu W-L, Yang L-C, Chung W-H, Chien Y-H, Hung C-F, et al. A Promoter Sequence Variant of ZNF750 Is Linked with Familial Psoriasis. J Invest Dermatol 2008; 128: 1662–1668.