

## Psoriatic Keratinocytes Express High Levels of Nerve Growth Factor

SIBA P. RAYCHAUDHURI, WEN-YUE JIANG and EUGENE M. FARBER

*Psoriasis Research Institute, Palo Alto, California, USA*

Many investigators have reported proliferation of terminal cutaneous nerves and upregulation of various neuropeptides (substance P, vasoactive intestinal polypeptide, calcitonin gene-related peptide) in psoriatic lesions. Nerve growth factor promotes growth of nerves and causes upregulation of neuropeptides like substance P and calcitonin gene-related peptide.

In this study we investigated the expression of nerve growth factor in psoriatic lesions, non-lesional psoriatic skin, lichen planus and normal control skin. Immunoperoxidase staining was applied on cryosections prepared from snap-frozen biopsy specimens. The primary antibody used was a polyclonal anti-NGF- $\beta$  antibody. Nerve growth factor was detected only in the keratinocytes. In psoriatic tissue the number of keratinocytes per square millimeter of epidermis positive for nerve growth factor was  $84.7 \pm 46.3$  compared to  $44.8 \pm 29.9$ ,  $18.9 \pm 11.8$  and  $7.5 \pm 16.9$ , respectively, in non-lesional psoriatic skin, normal skin and lichen planus.

Increased expression of nerve growth factor substantiates larger numbers of terminal cutaneous nerves and upregulations of substance P and calcitonin gene-related peptide in psoriatic lesions. In addition, nerve growth factor is mitogenic to keratinocytes, activates T-lymphocytes and can induce migration of inflammatory cellular infiltrates, histological features characteristic of psoriasis. **Key words:** neurogenic inflammation; NGF; pathogenesis; psoriasis.

(Accepted August 6, 1997.)

Acta Derm Venereol (Stockh) 1998; 78: 84–86.

Dr. Eugene M Farber, MD, Psoriasis Research Institute, 600, Town And Country Village, Palo Alto, CA 94301.

The pathogenesis of psoriasis remains largely unknown and is most likely a complex, multifactorial process. Clinical observations have suggested that the nervous system is involved in the disease process. The relationship between stress and the exacerbation of psoriasis is well documented (1–3). The striking symmetry of cutaneous lesions also raises the possibility that lesions may be related to the underlying peripheral innervation (4). Case reports have demonstrated remission of psoriasis in areas of traumatic denervation, with persistence of disease on the contralateral side (5). On the basis of these observations we proposed a role for the neuropeptides in the pathogenesis of psoriasis, suggesting that neurogenic inflammation triggered by exogenous and endogenous stimuli may be an underlying mechanism (4). Various investigators have explored this relatively new field of cutaneous neuroimmunology, and with the exceptions of a few reports (6, 7) others have reported an increased number of terminal cutaneous nerves along with upregulation of one or more of the neuropeptides such as substance P (SP), vasoactive intestinal polypeptide (VIP) and calcitonin gene-related peptide (CGRP) in psoriatic lesions (8–11).

The underlying causes for increased nerves and neuropep-

tides in the psoriatic skin have not been sufficiently addressed. As in other tissues, nerve growth factor (NGF) in the skin plays a role in regulating innervation (12). NGF also causes upregulation of neuropeptides (13). In this study, in order to understand the cause for the upregulation of neuropeptides and the hyperproliferation of terminal cutaneous nerves in psoriasis, we have investigated the expression of NGF in psoriatic plaques, non-lesional psoriatic skin, lichen planus and normal skin.

### MATERIAL AND METHODS

#### *Tissue preparation*

Biopsies were obtained from chronic psoriasis plaques ( $n=8$ ), non-lesional psoriatic skin ( $n=8$ ), skin of healthy individuals ( $n=5$ ) and lesional lichen planus skin ( $n=5$ ). The samples were snap-frozen with liquid nitrogen and stored in the refrigerator under  $-70^\circ\text{C}$ . The frozen samples were cut into 8- $\mu\text{m}$  cryosections. The sections were mounted on the glass slides and dried for 4 h at room temperature. Then the sections were immersed in 0.05 M, pH 3.0, citric acid buffered saline for 10 min and subsequently fixed with 4% formalin solution. The sections were washed and then sequentially blocked for endogenous biotin binding using the Vector blocking kit (Vector Laboratories, Burlingame, CA) and for endogenous peroxidase activity and non-specific antibody binding sites with 3% hydrogen peroxide and 1.5% normal goat serum (Vector Laboratories, Burlingame, CA) in 0.01M PBS.

#### *Immunohistochemistry staining*

The sections were first incubated for 24 h with 3  $\mu\text{g}/\text{ml}$  polyclonal anti-NGF- $\beta$  antibody (Chemicon International Inc., Temecula, CA) at room temperature. Standard immunoperoxidase techniques were applied for the rest of the procedures. Normal (non-immunized) rabbit serum (Sigma) in 1:20 dilution was used as a negative control. We also used NGF pre-absorption test of the NGF-antibody to rule out non-specific positivity.

### RESULTS

Positive staining was observed in the tissues stained only with the polyclonal antibody. Sections stained with the normal rabbit serum and the sections which were stained with the NGF antibody preabsorbed with NGF did not show any positive staining for NGF. All sections were examined by one investigator (WYJ), and independent confirmation of the numerical counting was performed by another investigator (SPRC). Cells in which staining could be appreciated without doubt were only considered to be positively stained. Cells which were slightly colored or where the positivity was doubtful were ignored. NGF was detected only in the keratinocytes. Tissues were examined for the presence of positively stained cells. Surface area of the epidermis was determined with the help of a reticle/grid (10  $\times$  10 mm with 1 mm<sup>2</sup> boxes; Microscoptics, Inc., Milford, MI) placed in the eye-piece. The number of cells positive for NGF in per square mm of epidermis was calculated by dividing the total number of

NGF-positive cells with the surface area. The data are described in Table I. In psoriatic tissues upper and mid epidermic keratinocytes expressed high levels of NGF (Fig. 1). The number of keratinocytes in per millimeter of epidermis stained for NGF was  $12.11 \pm 7.15$  in psoriatic tissues compared to  $2.55 \pm 1.71$ ,  $0.64 \pm 0.40$  and  $0.59 \pm 1.31$ , respectively, in non-lesional, normal skin and lichen planus ( $p < 0.01$ ). The differences in the NGF-positive keratinocyte numbers were more significant when keratinocytes/mm<sup>2</sup> were compared instead of keratinocytes/mm (Table I). NGF expression in both lesional ( $p < .01$ ) and non-lesional ( $p < .05$ ) psoriatic keratinocytes was significantly higher compared to the normal control skin and the lichen planus skin. The stratum corneum stained positively in both psoriasis and control skin. Positive staining of the stratum corneum was considered as a non-specific reaction.

## DISCUSSION

NGF is produced by the keratinocytes during embryonic development and during other circumstances which are not fully understood. In this study we have observed higher levels of NGF in the keratinocytes of the mid and upper epidermis of psoriatic tissues, compared to the controls (Fig. 1 A-C). As we know, psoriasis is a maturation disorder of the keratinocytes and therefore it is possible that immature keratinocytes at a certain phase of their cell cycle produce more NGF. Overexpression of NGF is known to induce nerve growth factor receptor (NGF-R) on the nerves (12). In a

separate study we have observed a marked upregulation of NGF-R in psoriatic lesions (14). Expression of larger amounts of NGF-R (14), along with an increased number of nerves (8), further substantiates an increased activity of NGF in psoriatic lesions.

It is worth noting that the expression of NGF is significantly higher in non-lesional psoriatic keratinocytes as well. In our study where we investigated for NGF-R expression we found similar results. We did not observe an upregulation of NGF in the keratinocytes of lichen planus cases. In our earlier study also we did not observe an increased expression of NGF-R in lichen planus (14). This suggests that the increased expression of NGF in the keratinocytes of lesional and non-lesional psoriatic tissue may not be a secondary event due to an inflammatory reaction.

NGF is mitogenic to keratinocytes (15, 16). NGF recruits mast cells and promotes their degranulation, both of which are early events in a developing lesion of psoriasis (17, 18). In addition NGF activates T lymphocytes and can recruit inflammatory cellular infiltrates (19-21). Thus, it is possible that expression of NGF is required before the influx of mast cells and lymphocytes, which in turn would initiate the inflammatory process of psoriasis.

Fantini et al. have earlier reported that NGF levels are higher in the tissue extracts from psoriatic lesions (22). This is the first direct evidence to show that the lesional psoriatic keratinocytes express high levels of NGF. Observations in this study that psoriatic keratinocytes produce a larger amount of

Table I. Expression of NGF in the keratinocytes of lesional and non-lesional psoriatic skin, lichen planus and normal skin

Biopsies	Numbers	No. of NGF-positive keratinocytes (KC) ( $\bar{x} \pm SD$ )	
		NGF <sup>+</sup> KC/mm	NGF <sup>+</sup> KC/mm <sup>2</sup>
Psoriatic skin	8	$12.11 \pm 7.15$	$84.68 \pm 46.35$
Non-lesional skin	8	$2.55 \pm 1.71$	$44.80 \pm 29.96$
Normal skin	5	$0.64 \pm 0.40$	$18.88 \pm 11.76$
Lichen planus	5	$0.59 \pm 1.31$	$7.54 \pm 16.86$

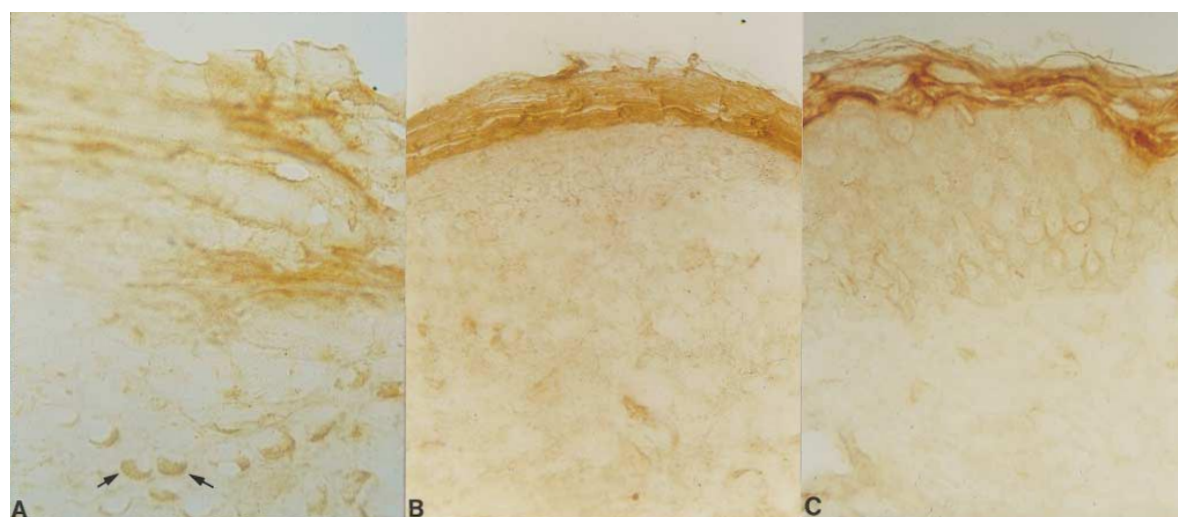


Fig. 1. Histological section of psoriatic plaque (A), demonstrating large amounts of NGF in the upper and mid epidermic keratinocytes compared to normal skin (B) and lichen planus (C). Arrows indicate the NGF-positive keratinocytes ( $\times 400$ ).

NGF compared to the controls constitute a significant finding. These observations further substantiate a role for the neurogenic inflammation in the pathogenesis of psoriasis and provide explanations for the following unanswered features of psoriasis: hyperproliferation of terminal cutaneous nerves, upregulation of neuropeptides (SP, CGRP) and disappearance of active psoriatic plaques at sites of anesthesia.

#### REFERENCES

- Farber EM, Nall ML. The natural history of psoriasis in 5600 patients. *Dermatologica* 1974; 148: 1–18.
- Gaston L, Lassonde M, Bernier-Buzzanga J, Hodgins S, Crombez JC. Psoriasis and stress: a prospective study. *J Am Acad Dermatol* 1987; 17: 82–86.
- Farber EM, Raychaudhuri SP. Concept of total care: a third dimension in the treatment of psoriasis. *Cutis* 1997; 59: 35–39.
- Farber EM, Nickoloff BJ, Recht B, Fraki JE. Stress, symmetry, and psoriasis: possible role of neuropeptides. *J Am Acad Dermatol* 1986; 14: 305–311.
- Raychaudhuri SP, Farber EM. Are sensory nerves essential for the pathogenesis of psoriasis. *J Am Acad Dermatol* 1993; 28: 448–449.
- Pincelli C, Fantini F, Romualdi P, Sevigani C, Lesa G, Benassi L, et al. Substance P is diminished and VIP is augmented in psoriatic lesions and these peptides exert disparate effects on the proliferation of cultured human keratinocytes. *J Invest Dermatol* 1992; 98: 421–427.
- Anand P, Springall DR, Blank MA, Sellu D, Polak JM, Bloom SR. Neuropeptides in skin disease: increased VIP in eczema and psoriasis but not axillary hyperhidrosis. *Br J Dermatol* 1991; 124: 547–549.
- Naukarinen A, Nickoloff BJ, Farber EM. Quantification of cutaneous sensory nerves and their substance P content in psoriasis. *J Invest Dermatol* 1989; 92: 126–129.
- Eedy DJ, Johnston CF, Shaw C, Buchanan KD. Neuropeptides in psoriasis: an immunocytochemical and radioimmunoassay study. *J Invest Dermatol* 1991; 96: 434–438.
- Al'Abadie MS, Senior HJ, Bleehen SS, Gawkrödger DJ. Neuropeptides and general neuronal marker in psoriasis—an immunohistochemical study. *Clin Exp Dermatol* 1995; 20(5): 384–389.
- Wallengren J, Ekman R, Sundler F. Occurrence and distribution of neuropeptides in human skin. An immunocytochemical and immunochemical study on normal skin blister fluid from inflamed skin. *Acta Derm Venereol (Stockh)* 1987; 67: 185–192.
- Wyatt S, Shooeter EM, Davies AM. Expression of the NGF receptor gene in sensory neurons and their cutaneous targets prior to and during innervation. *Neuron* 1990; 2: 421–427.
- Lindsay RM, Harmar AJ. Nerve growth factor regulates expression of neuropeptides genes in adult sensory neurons. *Nature* 1989; 337: 362–364.
- Farber EM, Chan J, Raychaudhuri SP, Smoller BR. Increased nerve growth factor receptor (NGF-R) in papillary dermis of lesional psoriatic skin: further evidence for a role of the sensory nervous systems in the pathogenesis of psoriasis. *Br J Dermatol* 1996; 135: 841.
- Wilkinson DI, Theeuwes MI, Farber EM. Nerve growth factor increases the mitogenicity of certain growth factors for cultured human keratinocytes: a comparison with epidermal growth factor. *Exper Dermatol* 1994; 3: 239–245.
- Pincelli C, Sevigani C, Manfredini R, Grande A, Fantini F, Bracci-Laudiero, et al. Expression and function of nerve growth factor and nerve growth factor receptor on cultured keratinocytes. *J Invest Dermatol* 1994; 103: 13–18.
- Aloe L, Levi-Mantalcini R. Mast cells increase in tissues of neonatal rats injected with the nerve growth factor. *Brain Res* 1977; 133: 358–366.
- Pearce FL, Thompson HL. Some characteristics of histamine secretion from rat peritoneal mast cells stimulated with nerve growth factor. *J Physiol* 1986; 372: 379–393.
- Bischoff SC, Dahinden CA. Effect of nerve growth factor on the release of inflammatory mediators by mature human basophils. *Blood* 1992; 79: 2662–2669.
- Otten U, Ehrhard P, Peck R. Nerve growth factor induces growth and differentiation of human B lymphocytes. *Proc Natl Acad Sci USA* 1989; 86: 10059–10063.
- Thorpe LW, Werrbach-Perez K, Perez-Polo JR. Effects of nerve growth factor on the expression of IL-2 receptors on cultured human lymphocytes. *Ann NY Acad Sci* 1987; 496: 310–311.
- Fantini F, Magnoni C, Brauci-Laudeis L, Pincelli C. Nerve growth factor is increased in psoriatic skin. *J Invest Dermatol* 1995; 105: 854–855.